MODELING OF MICROSCALE TRANSPORT IN BIOLOGICAL PROCESSES

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Modeling Cystic Fibrosis and Mucociliary Clearance

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5.1 MUCOCILIARY CLEARANCE AND CYSTIC FIBROSIS

This chapter considers the investigation of airway clearance efficiency and dysfunction using rheological measurements, modeling and simulations of mucus flows. The work is mainly dedicated to the respiratory epithelium dysfunction subsequently to the consumption of cilia-inhibiting drugs (e.g. nicotine) or viral and/or bacterial infections, and cystic fibrosis, although many respiratory diseases are associated with an altered mucus transport. The work presented here is a review based, amongst others, on Chatelin et al. (2015, 2016), Chatelin and Poncet (2013, 2014, 2016), Chatelin (2013).

The first part of this chapter focuses on the trachea–bronchial tree configuration, the respiratory epithelium, the dimensioning aspects and the cystic fibrosis pathology. After a factual dimensioning, the causes and effects of the cystic fibrosis are described. A mathematical model is finally built upon the nature of mucus and its surrounding epithelium environment.

The second part is devoted to the Newtonian modeling: efficient numerical methods are described. The algorithm is based on a penalization technique for the moving cilia-shaped...
geometry, with a one way coupling (the cell motion is prescribed). The asymptotic analysis and the well-posedness of the mathematical model are discussed. Specific numerical methods are introduced with their inherent features (convergence, computational time and memory use) as well as their use when mucociliary clearance is impaired and mucus viscosity rises.

The third part deals with non-Newtonian aspects. Measurements of cystic fibrosis patient mucus is described and summarized, leading to its rheological characterization. Mixing models for non-Newtonian rheology are based upon a continuous evolution from a low-viscosity Newtonian behavior, close to the respiratory epithelium wetted surface, to a strongly shear-thinning behavior in the mucus layer. Finally, an extension of the previous numerical method is set up in order to handle the non-Newtonian features, exhibiting non-reversibility of mucus motion and inefficiency of mucus transport at the cell scale for cystic fibrosis patient.

5.1.1 Airway Wall Environment and Mucociliary Clearance

The mucus in the human respiratory tract is a non-Newtonian fluid secreted by the respiratory epithelium. It protects the respiratory mucosa from dehydration and traps inhaled particles (dust, allergens, carcinogens, micro-organisms and inflammatory debris). It cleans airways, flowing from either the trachea–bronchial tree and larynx or nose and pharynx to the oesophagus entry, where it is swallowed (or expectorated), as displayed in Fig. 5.1. An adequate mucus mobility is necessary for a proper airway clearance. Otherwise pathogens can proliferate, and harmful agents can enter the respiratory mucosa.

The airway surface fluid is heterogeneous with a near-wall liquid layer containing mostly water and slightly polymerized mucins and a mucus layer in which completely polymerized mucins cross-link and build a meshwork, without discontinuity at the microscopic scale. In addition, the airway surface fluid moves due to the cyclic displacement of cilia. The moving fluid is thus associated with a multiscale problem both in time and space.

Respiratory ciliated epithelial cells have a collection of cilia on their luminal (wetted) surface. Over a given small airway segment, these cilia beat uniformly in the periciliary fluid (PCF) at a frequency of 4 to 20 Hz, quite different from that of the respiratory cycle (0.2–0.25 Hz). Mucus motion engendered by beating cilia of respiratory epithelial cells is influenced by its geometrical and rheological properties. The viscosity of the mucus film (thickness 10–100 µm) increases from its outer (close to the respiratory epithelium) to inner part (close to the core air flow).

Later in this section, a description is provided of the model that is used for numerical simulations of the motion of cilia immersed in a heterogeneous fluid with variable viscosity in the direction normal to the wall. The manifestation of cystic fibrosis is characterized by a stagnation of mucus, allowing a proliferation of pathogen agents and thus major infections, whose inflammation effects tend to destroy the epithelium ciliated cells, leading to sharper stagnation. It results that dangerous bacteria, such as *Pseudomonas aeruginosa* releasing pyocyanin, can then colonize the lung. Computing the resulting fluid displacement enables us to study the impact of different physical and biological parameters on the mucus transport at the scale
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of the cell, pointing out mechanisms involved in cystic fibrosis, the major pathological target of the present work.

5.1.2 The Cystic Fibrosis Pathology

Cystic fibrosis (CF) is a monogenic autosomal recessive disorder. This is the most frequent life-shortening genetic disease in the Caucasian population caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene on chromosome 7. These mutations lead to the production of a dysfunctional CFTR protein.

The prevalence of the disease varies according to location and ethnic background. CF affects more than 70,000 people worldwide: 30,000 in the USA, 6000 in France (Hoffman and Ramsey, 2013). It is observed in 1 infant out of 2500 in Europe and North America. The prevalence of cystic fibrosis in the United States of America is 0.797 per 10,000 inhabitants and 0.737 in the 27 European Union countries, with a highest value of 2.98 in the Republic of Ireland (Farrell, 2008).
More than 2000 mutations have been identified in the CFTR gene. Despite the allelic diversity in this genetic disease, about 85% of Caucasian people\textsuperscript{1} with CF pathology carry at least one copy of the CFTR gene mutation ΔF508 (F508\textsuperscript{del} CFTR channel variant, cf. Quon and Rowe, 2016). Approximately 4% of all Whites carry a single cystic fibrosis mutation. Carriers of a single cystic fibrosis mutation do not generally exhibit any phenotypic abnormality. Two mutations are required to be present for clinical manifestations of the disease.

In 1969, CF patients in industrialized countries had a mean survival of 14 years. Because of better understanding of relationships between CFTR gene defects and organ pathophysiology, life expectancy has improved substantially over the last 40 years. By 2010, the median predicted survival rate approached 40 years (Clancy and Jain, 2012).

The CFTR protein belongs to the large family of adenine nucleotide-binding cassette transporters consisting of two transmembrane domains, two nucleotide binding domains (NBDs) and a unique regulatory domain. This cAMP-regulated anion channel\textsuperscript{2} is expressed in the airway epithelium, submucosal glands and many other epithelial organs (e.g. biliary and pancreatic ducts) (Mall and Hartl, 2014). It transports anions (mainly chloride and bicarbonate) through the apical membrane of epithelial cells, thereby creating an osmotic gradient for fluid secretion; the active ion transport is coupled with water efflux. The basic defect causes an imbalance between secretion and absorption of the epithelial lining fluid rendering CF airway surfaces vulnerable to dehydration. The induced decrease in the thickness of the airway surface liquid is associated with an increased viscosity and thickening of airway secretions, an obstruction of airway lumens, a decreased ability to clear inhaled pathogens, and a dysregulated inflammation.

Studies in cultured primary airway epithelia from CF patients carrying a broad spectrum of CFTR mutations demonstrated that the airway surface liquid is depleted with a collapse of the periciliary layer and a mucociliary dysfunction (Mall and Hartl, 2014). In the following sections, the very different nature between periciliary fluid and mature mucus is investigated, and our results confirm the drop of the mucociliary clearance in the case of PCL collapsing.

Static airway mucus triggers inflammation of epithelial tissue and forms a nidus for bacterial infection. Defective CFTR-mediated bicarbonate transport leads to an acidic airway surface liquid that results in a decrease in antimicrobial activity (Haq et al., 2016). The epithelial sodium channel (ENaC) activity is upregulated in CF, which leads to sodium hyperabsorption and contributes to airway surface dehydration. ENaC activation occurs via various mechanisms including the cleavage of its subunit by endogenous proteases. Indeed, a damaging vicious circle of mucus retention, inflammation, infection is created (Hoffman and Ramsey, 2013). Many factors mediate the inflammatory response to chronic bacterial infection such as proinflammatory cytokines (IL-1β, IL-6, IL-8, GM-CSF, TNF-α). Sputum of CF contains elevated levels of neutrophils and hence exhibits a high concentration of neutrophil-derived proteases such as neutrophil elastase (Twigg et al., 2015).

Clinically, CF is a complex multi-organ disease that leads to a significant chronic sinopulmonary disease, pancreatic exocrine and endocrine impairments (diabetes), bowel disease,

\textsuperscript{1} In the general meaning of people from European extraction, including American white people and intermingling in former Europe.

\textsuperscript{2} cAMP stands for cyclic adenosine monophosphate, involved in the cAMP receptor proteins (CRP) binding tightly to a specific DNA.
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- Elevated sweat chloride, and male infertility (Zielenski, 2000). Chronic inflammation and infection may potentiate respiratory tissue injury, thereby altering progressively pulmonary function. The primary cause of death and long-term complications among people with CF is chronic bacterial infection in lungs (Yankaskas et al., 2004). Chronic obstructive lung disease evolves from an early onset of mucus plugging in small airways, chronic neutrophilic airway inflammation and bacterial infection (Zemanick et al., 2010).

- Cystic fibrosis patients battle life-long pulmonary infections. Chronic pulmonary infections contributes to a decline in forced expiratory volume in 1 s (FEV1) of approximately 2% per year. They result from a limited number of microbial pathogens, among which the most common species are *Haemophilus influenza*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and the *Burkholderia cepacia complex*. CF mucus is a specifically favorable niche for growth of *Pseudomonas aeruginosa* including glycosylation of airway mucins (Jeffries et al., 2016). There is sufficient evidence that eradication of early infection and prevention of chronic *Pseudomonas aeruginosa* infection is associated with clinical benefit (Doring et al., 2012).

- Until recently, medical treatments for CF exclusively targeted specific sequelae of the underlying pulmonary disease. For example, intravenous or inhaled antibiotics are administered to limit bacterial infection while macrolides or ibuprofen can be used to reduce inflammation (Doring et al., 2012). Restoring the impaired clearance of airway secretions in order to optimize micro-organisms removal from airways is one of the major goals of therapy in patients with CF (Kurbatova et al., 2015).

- Current therapies are varied and include: (i) airway clearance to mobilize bronchopulmonary secretions and interrupt the vicious cycle of inflammation and infection (McShane et al., 2013), (ii) inhaled dornase which, while it does break down DNA and decrease mucus viscosity, has recently been shown to require a peripheral small airway deposit (Kurbatova et al., 2015), (iii) hypertonic saline which may improve the airway surface hydration (Hoffman and Ramsey, 2013), and (iv) mannitol that can enhance the airway mucociliary clearance (Aitken et al., 2012; Bilton et al., 2011).

- New pharmacotherapy approaches focus on the development of CFTR modulators. These CFTR-targeted drugs are designed to rescue mutant CFTR function (Mall and Hartl, 2014). Correctors relieve protein misprocessing due to the most common mutation ΔF508, as they facilitate proper maturation and delivery to the plasma membrane. Potentiators are CFTR modulators that increase the time CFTR spends in the open channel configuration and hence chloride transport. They restore airway surface hydration and mucus clearance. Combined therapies (potentiators and correctors) are now available for patients (Quon and Rowe, 2016).

- Alternative therapeutic approaches to improve airway surface hydration include stimulation or inhibition of other channels and transporters involved in apical ion transport to obviate the abnormal ion transport, solute transfer, pH and airway surface liquid volume generated by dysfunctional CFTRs. Given hyperactivity of ENaC in CF, inhibitors of the ENaC channel or activators of alternative Cl⁻ channels (CaCC) have recently been investigated as potential therapies (Haq et al., 2016).

5.1.3 Mathematical Modeling of Lung Wall Environment

Since the 1970s and the paper of Blake (1972), the mucociliary clearance has been frequently studied using mathematical modeling combined with numerical and analytic com-
putations – see Smith et al. (2008) for a review. Since the publication of this work (2008), several researchers published new researches using 2D modeling (Smith et al., 2007b; Lukens et al., 2010; Montenegro-Johnson et al., 2013; Lee et al., 2011; Jayathilake et al., 2015; Sedaghat et al., 2016) or 3D modeling (Jayathilake et al., 2012; Simons et al., 2015; Smith et al., 2007a; Li et al., 2016; Ding et al., 2014; Ding and Kanso, 2015; Guo and Kanso, 2016a, 2016b).

In these papers the airway surface liquid is often considered to have a two-layer structure with constant viscosity in each layer (Lee et al., 2011; Jayathilake et al., 2015; Li et al., 2016). Alternatively, other studies assume a constant viscosity airway surface liquid composed only with one fluid layer (Jayathilake et al., 2012; Simons et al., 2015). The two layer model has also been combined with rheological behaviors in the upper mucus layer, using a shear-thinning modeling (Montenegro-Johnson et al., 2013; Craster and Matar, 2000) or a viscoelastic modeling (Mitran, 2007; Sedaghat et al., 2016; Guo and Kanso, 2016b).

Regarding numerical analysis, the mucociliary clearance has been computed using various techniques. As the flow modeling leads to a Stokes problem, the Stokeslets method (Cortez et al., 2005) has been used by several authors (Smith et al., 2007a, 2007b; Lukens et al., 2010; Montenegro-Johnson et al., 2013; Guo and Kanso, 2016a). The main drawbacks of this method include the computational cost which grows with the number of immersed cilia and becomes prohibitive in 3D, as well as the limitation to constant viscosity flows avoiding the computation of non-linearities such as rheology.

Finite volumes have also been used: a complete discretization coupled to a finite element beam model for cilia has been published (Mitran, 2007), taking into account the fluid–structure interaction. The flow was successfully computed but this technique requires a large amount of computational resources to solve resulting linear systems. The immersed boundary method, introduced by Peskin (2003) was also used in a finite differences context for a parametric study. More recently the immersed boundary method was coupled with Lattice Boltzmann techniques in order to take into account the two-layer structure of the airway surface liquid (Li et al., 2016; Sedaghat et al., 2016; Guo and Kanso, 2016a, 2016b).

Because the characteristic dimensions of the mucus flow are very small (the mucus film is 10–40 μm height and cilia are 5–8 μm long), the inertia terms of the Navier–Stokes equations are negligible, and the conservation of momentum can be represented by the linearized Stokes equations. In fact, at this scale inertia terms vanish in the Navier–Stokes equations. To simulate the mucociliary clearance, it is important to take into account the cilia motion which propels the airway surface liquid by means of the fluid–structure interaction. In the following model description, the fluid–structure interaction is assumed to be “one way”: the force exerted by the fluid on the solid is not computed. In fact, this remains an open problem from a numerical point of view, in particular an eventual beating adaptation (when the airway surface liquid becomes more viscous, for example) has never been studied numerically and, to our knowledge, only one experimental work was reported (Hill et al., 2010).

The existence of an interface between the periciliary fluid and mucus is debated (Button et al., 2012; Ehre et al., 2014). In fact, polymerized mucins are released in the periciliary fluid, where they become progressively hydrated during their crossing of the PCL. Mucins accumulate at a given distance normal to the wall, giving rise to a mucin-concentrated phase, the mucus. In this region, mucin polymers interweave and eventually cross-link. The whole set of transformations undergone by mucins during their transport through the ASL is referred to as maturation in the present text.
These mucins are primarily responsible for the mucus structure and its highly non-Newtonian behavior. Near the epithilium the mucin concentration is very low and the airway fluid behaves as a Newtonian fluid, while further away from the epithilium, the presence of high mucin concentrations results in non-Newtonian shear thinning behavior (as described on Figure 5.2). In order to represent the airway fluid, and particularly its viscosity, as a continuum instead of a two layer fluid, a variable $\alpha$ is introduced that is a function of position that varies in a smooth sigmoidal manner between 0 and 1. This function is defined in greater detail later in this section.

This means that the viscosity can be seen as an \textit{ad-hoc} function of $\alpha$: Either $\mu = \Phi(\alpha)$ in the Newtonian case, detailed in Section 5.2, or $\mu = \Phi(u, \alpha)$ in the non-Newtonian case, typically

$$\mu = k(\alpha)(2D : D)^{(n(\alpha) - 1)/2}$$

for power-law fluids, or a Carreau law for the fluids whose rheology exhibits thresholds, where $D$ is the strain tensor defined below, detailed in Section 5.3. Then starting from an initial distribution, this variable $\alpha$ which characterizes mucins evolution, is transported and diffused by the mucus flow which depends on the viscosity. Hence $\alpha$ is the solution to a convection–diffusion equation and it is the natural to assume that the viscosity is bounded: $\mu_0 \leq \Phi(\alpha) \leq \mu_1$.

The mathematical problem which has to be solved is the following:

\begin{equation}
\begin{cases}
  u = \bar{u} & \text{inside cilia}, \\
  -\text{div}(2\mu \mathbf{D}(u)) = f - \nabla p & \text{inside fluid}, \\
  \text{div} u = 0 & \text{inside fluid}, \\
  \partial_t \alpha + u \cdot \nabla \alpha - \eta \Delta \alpha = 0 & \text{everywhere in computational domain}, \\
  \mu = \Phi(\alpha) \text{ or } \mu = \Phi(\alpha, u) & \text{everywhere in computational domain},
\end{cases}
\end{equation}

where $p$ is the pressure, $\mu$ is the fluid viscosity, $f$ are the external forces and $\mathbf{D}(u) = (\nabla u + (\nabla u)^T)/2$ is the strain tensor and $\bar{u}$ is the cilia velocity (see computational details in Section 5.2.2.1). With initial and boundary conditions yet to be set.

To take into account the motion of immersed beating cilia, the Penalization method is used (Angot et al., 1999). It consists in adding an extra term to the conservation of momentum to solve a unique equation in all the domain, without any distinction between the cilia region $B(t)$ and the fluid region $\Omega \setminus B(t)$ (assuming that $\Omega$ is the computational domain). This extra Penalization term forces the velocity $u$ to be equal to a prescribed solid velocity $\bar{u}$ in the solid region $B(t)$. It transforms the previous problem into the Penalized Stokes problem or the Stokes–Brinkman problem coupled to the same convection–diffusion equation generating a non-linear dynamics:

\begin{equation}
\begin{cases}
  -\text{div}(2\mu^\varepsilon \mathbf{D}(u^\varepsilon)) + \frac{1_{B(t)}}{\varepsilon} (u^\varepsilon - \bar{u}) = f - \nabla p^\varepsilon & \text{in } \Omega, \\
  \text{div} u^\varepsilon = 0 & \text{in } \Omega, \\
  \partial_t \alpha^\varepsilon + u^\varepsilon \cdot \nabla \alpha^\varepsilon - \eta \Delta \alpha^\varepsilon = 0 & \text{in } \Omega, \\
  \mu^\varepsilon = \Phi(\alpha^\varepsilon) \text{ or } \mu^\varepsilon = \Phi(\alpha^\varepsilon, u^\varepsilon) & \text{in } \Omega,
\end{cases}
\end{equation}

with $1_{B(t)}$ the characteristic function of the solid domain and $\varepsilon$ the penalty parameter.
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FIGURE 5.2 Sketch of the mucus flow on the bronchial wall: the non-homogeneous mucus, which exhibits a smooth transition between PCL and maturated mucus, is propelled by beating ciliated epithelium cells, from Chatelin and Poncet (2016).

This coupled non-linear problem will be solved using dedicated algorithms in order to get powerful 3D simulations and to allow parametric investigations. As detailed in Section 5.2.2, these algorithms are based on fast solvers on regular Cartesian grids, so that the computational domain $\Omega$ is assumed to be a rectangular cuboid.

The Stokes problem is completed with a no slip boundary condition at the epithelium surface (where the periciliary fluid is present), and a free slip boundary condition at the air–mucus interface (which is assumed to be flat due to a strong surface tension effect, Enault et al., 2010; Chatelin et al., 2016) and periodic boundary conditions in the other (proximal and transverse) directions. In the following, let $\Gamma_0$ and $\Gamma_L$ respectively denote the upper and lower boundaries of the rectangular cuboid computational domain. Hence $\Gamma_0$ and $\Gamma_L$ respectively are the boundary regions where the homogeneous Dirichlet and homogeneous Neumann boundary conditions are imposed.

For the convection–diffusion equation, homogeneous Neumann boundary conditions are set at both epithelium–periciliary–fluid-layer and air–mucus interfaces (meaning on $\Gamma_0 \cup \Gamma_L$) and periodic boundary conditions in other directions.

Moreover, an initial condition is provided for the mucins ratio $\alpha$. A sigmoid profile is used to model the cross-linked mucins ratio which continuously varies from 0 in the deeper periciliary fluid layer to 1 in the upper mucus layer:

$$\alpha(x, y, z, t = 0) = \frac{\arctan(y(z - \delta)) - \arctan(-y\delta)}{\arctan(y(H_z - \delta)) - \arctan(-y\delta)}$$

where $\delta$ is the ratio between the mucus layer height and the periciliary fluid layer height, and $z$ denotes the coordinate perpendicular to the epithelium designating position in the fluid. A depiction of Eq. (5.3) is presented in Fig. 5.3.

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A Newtonian fluid behavior means that the viscosity $\mu$ only depends on $\alpha$, and is independent of $u$. In Eqs. (5.1) and (5.2), the case $\mu = \Phi(\alpha)$ is considered. This section is divided into
three parts: firstly, the well-posedness of Eq. (5.2) is demonstrated; secondly, the numerical algorithms dedicated to the computation of this problem are presented; and finally, several computations are presented as well as phenomenological analysis.

5.2.1 Mathematical Analysis

In order to perform a mathematical analysis of the problem, we consider first a regularization of the domain \( B(t) \) described in Fig. 5.4. From a technical point of view, this allows using trace and lifting theorems in these two domains and justifying the pertinence of the Penalization method. For the sake of simplicity, the notation \( B(t) \) refers to this regularized domain in the whole Section 5.2.1.

Let \( T > 0 \) be a final time (not necessarily a cilia period). Let \( \Gamma = [0, L_1] \times [0, L_2] \) and \( \Omega = \Gamma \times [0, L] \) be the domain of study. We denote by \( \Gamma_0 = \Gamma \times \{0\} \) and \( \Gamma_L = \Gamma \times \{L\} \) the lower and upper boundaries of \( \Omega \).

Since the domain \( B \) depends on the time parameter \( t \in [0, T] \), we need to track this dependency in the following. We let

\[
O = [0, T] \times \Omega \quad \text{be the whole domain in time and space coordinates,}
\]

\[
O_p = \{(t, x) \in [0, T] \times \Omega, \forall t \in [0, T], x \in B(t)\} \quad \text{be the penalized domain,}
\]

\[
\Sigma = \{(t, x) \in [0, T] \times \Omega, \forall t \in [0, T], x \in \partial B(t) \setminus \partial \Omega\} \quad \text{be the inner boundary of } O_p.
\]

Problem (5.1) is rewritten in these spaces as

\[
\begin{align*}
\mu & = \ddot{u} & \text{in } O_p, \\
-\text{div}(2\mu D(u)) &= f - \nabla p & \text{in } O'_p, \\
\text{div} u &= 0 & \text{in } O, \\
\partial_t \alpha + u \cdot \nabla \alpha - \eta \Delta \alpha &= 0 & \text{in } O, \\
\mu &= \Phi(\alpha) & \text{in } O.
\end{align*}
\]
with the initial and boundary conditions described previously:

\[\begin{align*}
&\begin{cases}
  u = 0 & \text{on } [0, T] \times \Gamma_0, \\
  u = \bar{u} & \text{on } \Sigma, \\
  u \cdot v = \frac{\partial u_1}{\partial v} = \frac{\partial u_2}{\partial v} = 0 & \text{on } [0, T] \times \Gamma_L, \\
  u \text{ is periodic} & \text{on } [0, T] \times \partial \Gamma \times [0, L], \\
  \frac{\partial \alpha}{\partial v} = 0 & \text{on } [0, T] \times (\Gamma_0 \cup \Gamma_L), \\
  \alpha \text{ is periodic} & \text{on } [0, T] \times \partial \Gamma \times [0, L], \\
  \alpha(0, \cdot) = \alpha_0 & \text{in } \Omega,
\end{cases}
\end{align*}\]  

(5.5)

where \(\eta > 0\) is the diffusion coefficient, \(\Phi \in W^{3,\infty}(\mathbb{R})\) and there exists a real number \(\mu_0 > 0\) such that for all \(\alpha \in \mathbb{R}, \mu_0 \leq \phi(\alpha)\).

According to the model, the penalized problem is rewritten as

\[\begin{align*}
&\begin{cases}
  -\text{div}(2\epsilon \mu^\varepsilon D(u^\varepsilon)) + \frac{1}{\epsilon} \Phi(u^\varepsilon - \bar{u}) = f - \nabla p^\varepsilon & \text{in } O, \\
  \text{div } u^\varepsilon = 0 & \text{in } O, \\
  \delta_1 \alpha^\varepsilon + u^\varepsilon \cdot \nabla \alpha^\varepsilon - \eta \Delta \alpha^\varepsilon = 0 & \text{in } O, \\
  \mu^\varepsilon = \Phi(\alpha^\varepsilon) & \text{in } O.
\end{cases}
\end{align*}\]  

(5.6)
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with its initial and boundary conditions coming straight from Eqs. (5.5):

\[
\begin{align*}
    u^\varepsilon &= 0 & \text{on } [0, T] \times \Gamma_0, \\
    u^\varepsilon \cdot v &= \frac{\partial u^\varepsilon}{\partial v} = 0 & \text{on } [0, T] \times \Gamma_L, \\
    u^\varepsilon &\text{ is periodic} & \text{on } [0, T] \times \partial \Gamma \times [0, L], \\
    \frac{\partial u^\varepsilon}{\partial v} &= 0 & \text{on } [0, T] \times (\Gamma_0 \cup \Gamma_L), \\
    a^\varepsilon &\text{ is periodic} & \text{on } [0, T] \times \partial \Gamma \times [0, L], \\
    a^\varepsilon(0, \cdot) &= a_0 & \text{in } \Omega.
\end{align*}
\]

(5.7)

Finally, the following functional spaces are introduced:

**Definition 1.** Let \( F(\Omega; X) \) be the set of functions defined on \( \Omega \) with values in \( X \) and

\[
\begin{align*}
    L^2(\Omega) &= \{ v \in L^2(\Omega), \int_\Omega v \, dx = 0 \}, \\
    V(\Omega) &= \{ v \in H^1(\Omega), \text{div} v = 0 \text{ on } \Omega, u = 0 \text{ on } \Gamma_0, \\
    &\quad \text{on } [0, T] \times \Gamma_L, u \text{ has periodic conditions on } \partial \Gamma \times [0, L] \}\}
\]

with the classical \( H^1 \)-norm. Thanks to the generalized Poincaré's inequality (see Boyer and Fabrie, 2005, Section III.6), there exist two constants depending only on \( \Omega \) such that

\[
c_1 \| \nabla v \|_{L^2(\Omega)} \leq \| v \|_{H^1(\Omega)} \leq c_2 \| \nabla v \|_{L^2(\Omega)}.
\]

The following existence and uniqueness results require some regularity assumptions on the obstacle’s time evolution (H1)–(H2) and on the velocity \( \bar{u} \) inside the obstacle (H3):

**Hypothesis 1 (H1).**

- Domain \( B \) is an open set in \( \Omega \) having the uniform \( C^2 \)-regularity property (see Adams and Fournier, 2003, Section 4 for the definition) up to the boundary of \( \Omega \) in order to obtain a periodic smooth domain,
- \( \exists (l, l') \in (\mathbb{R}^+)^2 \) such that \( 0 < l < l' < L \) and \( \Gamma \times [0, l] \subset B \subset \Gamma \times [0, l'] \), i.e. we assume that the bronchial wall is the fixed domain \( \Gamma \times [0, l] \) to which cilia are attached and that the cilia never reach the top of the domain \( \Omega \),
- \( \Omega \setminus B \) is an open set in \( \Omega \) having the uniform \( C^2 \)-regularity property,
- \( \Omega \setminus B \) is simply connected.

**Hypothesis 2 (H2).**

- There exists a function \( \Psi \in C^0([0, T]; C^2(\Omega)) \) such that for all \( t \in [0, T] \) \( \Psi(t)(0, l) \times \Gamma = \text{Id}_{[0, L] \times \Gamma}, \Psi(t)(0, l') \times \Gamma \subset [0, l'] \times \Gamma \) and \( \Psi(t) \) has periodic boundary conditions on \( [0, L] \times \partial \Gamma \),
- \( \forall t \in [0, T] \Psi(t) \) is a \( C^2 \)-diffeomorphism on \( \Omega \),
- \( \forall t \in [0, T] B(t) = \Psi(t, B) \).

**Remark.** Here the assumption has been made that the bronchial wall never moves and that the cilia never reaches the top of the domain. Additionally, \( \Omega \setminus B(t) = \Psi(t, \Omega \setminus B) \).
Hypothesis 3 (H3).
- $\tilde{u} \in C^0([0, T]; H^1(B(\cdot)))$,
- $\forall t \in [0, T], \text{div } \tilde{u}(t, \cdot) = 0$ in $B(t)$,
- $\forall t \in [0, T], \tilde{u}(t, \cdot)$ fulfills
  $$\forall t \in [0, T] \quad \tilde{u}(t, \cdot)|_{\Gamma_0} = 0,$$
- $\forall t \in [0, T], \tilde{u}(t, \cdot)$ has periodic boundary conditions on $\partial B(t) \cap (\partial \Gamma \times [0, L])$.

Definition 2. $(u^\varepsilon, p^\varepsilon, \alpha^\varepsilon)$ is a weak solution to (5.6)–(5.7) if it fulfills
- $u^\varepsilon \in L^q(0, T; V(\Omega)), \ p^\varepsilon \in L^q(0, T; L^2_0(\Omega))$ with $2 \leq q \leq +\infty, \ \alpha^\varepsilon \in L^\infty(0, T; H^2(\Omega)) \cap L^2(0, T; H^1(\Omega)) \cap H^1(0, T; H^1(\Omega))$,
- For all $v \in C_c(0, T; V(\Omega))$,
  $$\iint_{(0,T) \times \Omega} 2 \mu^\varepsilon D(u^\varepsilon) : D(v) \, dx \, dt + \frac{1}{\varepsilon} \iint_{\Omega} (u^\varepsilon - \tilde{u}) \cdot v \, dx \, dt = \iint_{(0,T) \times \Omega} f \cdot v \, dx \, dt, \quad (5.8)$$
- $\alpha^\varepsilon$ is the solution to
  $$\begin{cases}
  \partial_t u^\varepsilon + u^\varepsilon \cdot \nabla \alpha^\varepsilon - \eta \Delta \alpha^\varepsilon = 0 & \text{in } \Omega, \\
  \frac{\partial u^\varepsilon}{\partial t} = 0 & \text{on } \Gamma_0 \cup \Gamma_L, \\
  \alpha^\varepsilon \text{ has periodic conditions} & \text{on } \partial \Gamma \times [0, L], \\
  \alpha^\varepsilon(0, \cdot) = \alpha_0 & \text{in } \Omega,
  \end{cases}
  \quad (5.9)$$
- $\mu^\varepsilon = \Phi(\alpha^\varepsilon)$.

Theorem 1. Let $0 < \mu_0$ and $\varepsilon > 0$. Let $\alpha_0 \in H^2(\Omega)$ and $\Phi \in W^{2, \infty}(\mathbb{R})$ such that $\forall x \in \mathbb{R}, \ \mu_0 \leq \Phi(x)$.
We moreover assume (H1), (H2), (H3) and that for all $T > 0, \ f \in L^\infty(0, T; L^2(\Omega))$. There exists then a unique weak solution $(u^\varepsilon, p^\varepsilon, \alpha^\varepsilon)$ to (5.6) such that
$$\begin{cases}
  \alpha^\varepsilon \in L^\infty(0, T; H^2(\Omega)) \cap L^2(0, T; H^3(\Omega)) \cap H^1(0, T; H^1(\Omega)), \\
  u^\varepsilon \in L^q(0, T; V(\Omega)) \quad \text{for all } 2 \leq q \leq +\infty, \\
  p^\varepsilon \in L^q(0, T; L^2_0(\Omega)) \quad \text{for all } 2 \leq q \leq +\infty.
\end{cases}$$

Moreover, there exist some constants independent of $\varepsilon$ such that $u^\varepsilon$ fulfills
$$\begin{align*}
\mu_0 \| D(u^\varepsilon) \|^2_{L^2([0,T];L^2(\Omega))} + \frac{1}{\varepsilon} \| u^\varepsilon - \tilde{u} \|^2_{L^\infty([0,T];L^2(B(\cdot)))} & \leq C \| f \|^2_{L^\infty([0,T];L^2(\Omega))} \| \tilde{u} \|^2_{L^\infty([0,T];H^2(B(\cdot)))} \\
& \quad + C'( \| f \|^2_{L^\infty([0,T];L^2(\Omega))} + \| \Phi \|^2_{L^\infty(\mathbb{R})} \| \tilde{u} \|^2_{L^\infty([0,T];H^2(B(\cdot)))} ), \\
\mu_0 \| D(u^\varepsilon) \|^2_{L^\infty([0,T];L^2(\Omega))} + \frac{1}{\varepsilon} \| u^\varepsilon - \tilde{u} \|^2_{L^2([0,T];L^2(B(\cdot)))} & \leq C \| f \|^2_{L^2([0,T];L^2(\Omega))} \| \tilde{u} \|^2_{L^\infty([0,T];H^2(B(\cdot)))} \\
& \quad + C'( \| f \|^2_{L^\infty([0,T];L^2(\Omega))} + \| \Phi \|^2_{L^\infty(\mathbb{R})} \| \tilde{u} \|^2_{L^\infty([0,T];H^2(B(\cdot)))} ).
\end{align*}$$
Definition 3. \((u, p, \alpha)\) is a weak solution to (5.4) with the conditions (5.5) if it fulfills

- For all \(2 \leq q \leq +\infty\), \(u \in L^q(0, T; V(\Omega))\), \(p \in L^q(0, T; L^2_0(\Omega \setminus \bar{B}(\cdot)))\) and \(\alpha \in L^\infty(0, T; H^2(\Omega)) \cap L^2(0, T; H^3(\Omega)) \cap \bar{H}^1(0, T; H^1(\Omega))\),
- \(u = \bar{u}\) in \(O_\rho\),
- \((u, p) \in L^q(0, T; H^1(\Omega \setminus \bar{B}(\cdot))) \times L^q(0, T; L^2_0(\Omega \setminus \bar{B}(\cdot)))\), with \(2 \leq q \leq +\infty\), is the solution to

\[
\begin{align*}
-\text{div}(2\mu D(u)) &= f - \nabla p \quad \text{in } \Omega, \\
\text{div } u &= 0 \quad \text{in } \Omega, \\
u \cdot v &= \frac{\partial u_1}{\partial v} = \frac{\partial u_2}{\partial v} = 0 \quad \text{on } [0, T] \times I_L, \\
u^n \text{ has periodic conditions on } \partial \Gamma \times [0, L],
\end{align*}
\]

- \(\alpha\) is the solution to (5.9),
- \(\mu = \Phi(\alpha)\).

Theorem 2. Under the assumptions of Theorem 1 and with the extra assumption \(\Phi \in W^{3,\infty}(\mathbb{R})\), there exists a unique weak solution \((u, p, \alpha)\) to (5.4)–(5.5). Moreover, the weak solution \((u^n, p^n, \alpha^n)\) of (5.6) converges weakly towards \((u, p, \alpha)\) as \(\varepsilon\) goes to zero.

We now present the sketches of the proofs of Theorems 1 and 2 and highlight the key points in getting these theorems.

5.2.1.1 Sketch of the Proof of Theorem 1

To prove the existence of a solution to this coupled non-linear problem, introduce the sequence \((\alpha^n, u^n, p^n)\) defined by:

1. \(\alpha^0 = \alpha_0\),
2. \(\mu^n = \Phi(\alpha^n)\),
3. \((u^n, p^n)\) is the solution to

\[
\begin{align*}
-\text{div}(2\mu^n D(u^n)) + \frac{1}{\varepsilon} (u^n - \bar{u}) &= f - \nabla p^n \quad \text{in } \Omega, \\
\text{div } u^n &= 0 \quad \text{in } \Omega, \\
u^n &= 0 \quad \text{on } I_0, \\
u \cdot v &= \frac{\partial u_1}{\partial v} = \frac{\partial u_2}{\partial v} = 0 \quad \text{on } I_L, \\
u^n \text{ has periodic conditions on } \partial \Gamma \times [0, L],
\end{align*}
\]

4. \(\alpha^{n+1}\) is the solution to

\[
\begin{align*}
\partial_t \alpha^{n+1} + u^n \cdot \nabla \alpha^{n+1} - \eta \Delta \alpha^{n+1} &= 0 \quad \text{in } \Omega, \\
\frac{\partial \alpha^{n+1}}{\partial \nu} &= 0 \quad \text{on } I_0 \cup I_L, \\
\alpha^{n+1} \text{ has periodic conditions} \quad \text{on } \partial \Gamma \times [0, L], \\
\alpha^{n+1}(0, \cdot) &= \alpha_0 \quad \text{in } \Omega.
\end{align*}
\]

We let \(\alpha^{n+1} = G(\alpha^n)\).
The study of this recurring sequence leads us to the study of the convection–diffusion equation, on the one side, and the penalized Stokes equation, on the other side.

**Theorem 3.** Let $\eta > 0$ and $u \in L^q(0, T; V(\Omega))$ for all $2 \leq q \leq +\infty$. Let $a_0 \in \mathbb{H}^1(\Omega)$. There exists a unique solution $\alpha \in L^\infty(0, T; H^1(\Omega)) \cap L^2(0, T; \mathbb{H}^2(\Omega)) \cap H^1(0, T; L^2(\Omega))$ to the problem (5.9).

Moreover, if $a_0 \in \mathbb{H}^2(\Omega)$ then $\alpha \in L^\infty(0, T; \mathbb{H}^2(\Omega)) \cap L^2(0, T; \mathbb{H}^3(\Omega)) \cap H^1(0, T; H^1(\Omega))$, and there exist some constants $C$ and $C'$ depending only on $\Omega$ and $\eta$ such that for all $t \in [0, T]$: 

\[
\|\alpha(t, \cdot)\|_{\mathbb{H}^2(\Omega)} \leq C\|a_0\|_{\mathbb{H}^2(\Omega)} \exp\left( C' \int_0^t \left( 1 + \|u(s, \cdot)\|_{H^1(\Omega)}^2 + \|u(s, \cdot)\|_{H^1(\Omega)}^4 + \|u(s, \cdot)\|_{H^1(\Omega)}^8 \right) ds \),
\]

where $\|\alpha\|_{L^2(0, T; \mathbb{H}^1(\Omega))}$ and $\|\alpha\|_{L^2(0, T; \mathbb{H}^2(\Omega))}$.

**Proof.** A classical Galerkin decomposition is performed, and energy estimates are made in order to obtain this result (see also Dautray and Lions, 2000, Vol. 5, Section XVIII).

**Theorem 4.** Assume that hypothesis (H1), (H2) and (H3) are satisfied. Let $\mu_0 > 0$, $\varepsilon > 0$ and $T > 0$. Let $\mu \in L^\infty([0, T] \times \Omega)$ such that $\mu_0 \leq \mu(t, x)$ a.e. in $[0, T] \times \Omega$. Let $f \in L^\infty(0, T; L^2(\Omega))$. There exists then a unique weak solution $(u, p)$ to the problem

\[
\begin{align*}
-\text{div}(2\mu D(u)) + \frac{1}{\varepsilon}(u - \bar{u}) = f - \nabla p & \quad \text{in } [0, T] \times \Omega, \\
\text{div} u = 0 & \quad \text{in } [0, T] \times \Omega, \\
\mu \geq 0 & \quad \text{on } [0, T] \times \Gamma_0, \\
\frac{\partial u_1}{\partial\nu} = \frac{\partial u_2}{\partial\nu} = 0 & \quad \text{on } [0, T] \times \Gamma_L, \\
u \text{ has periodic conditions} & \quad \text{on } [0, T] \times \partial \Gamma \times [0, L],
\end{align*}
\]

which fulfills

\[
(u, p) \in L^q([0, T], V(\Omega)) \times L^q\left([0, T]; L^2_0(\Omega)\right)
\]

for all $2 \leq q \leq +\infty$.

Additionally, the following inequalities hold:

\[
\begin{align*}
\mu_0\|D(u)\|_{L^\infty([0, T]; L^2(\Omega))}^2 & + \frac{1}{\varepsilon}\|u - \bar{u}\|_{L^\infty([0, T]; L^2(B(\cdot)))}^2 \\
& \leq C\|f\|_{L^\infty([0, T]; L^2(\Omega))}\|\bar{u}\|_{L^\infty([0, T]; H^1(\Omega))}^2 + C'\left(\|f\|_{L^\infty([0, T]; L^2(\Omega))}^2 + \|\mu\|_{L^\infty([0, T]; L^2(\Omega))}^2\|\bar{u}\|_{L^\infty([0, T]; H^1(\Omega))}^2\right),
\end{align*}
\]

\[
\begin{align*}
\mu_0\|D(u)\|_{L^2([0, T]; L^2(\Omega))}^2 & + \frac{1}{\varepsilon}\|u - \bar{u}\|_{L^2([0, T]; L^2(B(\cdot)))}^2 \\
& \leq C\|f\|_{L^2([0, T]; L^2(\Omega))}\|\bar{u}\|_{L^\infty([0, T]; H^1(\Omega))}^2 + C'\left(\|f\|_{L^2([0, T]; L^2(\Omega))}^2 + \|\mu\|_{L^2([0, T]; L^\infty(\Omega))}^2\|\bar{u}\|_{L^\infty([0, T]; H^1(\Omega))}^2\right).
\end{align*}
\]
Moreover, if \( \mu \in L^q(0, T; W^{1, \infty}(\Omega)) \) with \( 2 \leq q \leq +\infty \), then
\[
(u, p) \in L^q \left( [0, T], H^2(\Omega) \right) \times L^q \left( [0, T]; \mathbb{H}^1(\Omega) \right),
\]
and there exists a constant \( C \) depending on \( \Omega \) such that
\[
\|u\|_{L^q([0, T]; H^2(\Omega))} + \left\| \frac{p}{\mu} \right\|_{L^q([0, T]; \mathbb{H}^1(\Omega))} \leq C \|g\|_{L^q([0, T]; L^2(\Omega))},
\]
where \( g = \frac{1}{\mu} \left( 2D(u) \nabla \mu + f - \frac{1_{B(0)}}{\epsilon} (u - \bar{u}) + p \frac{\nabla \mu}{\mu} \right) \in L^q([0, T]; L^2(\Omega)) \).

**Proof.** The existence and uniqueness may be obtained classically thanks to Lax–Milgram theorem. The regularity is due to classical Stokes regularity results (see, for example, Boyer and Fabrie, 2005) thanks to the following rewriting of the equation:
\[
- \text{div} \, 2D(u) = \frac{1}{\mu} \left( -2D(u) \nabla \mu - \frac{1_{B(0)}}{\epsilon} (u - \bar{u}) + f - \nabla p \right).
\]
The \( \epsilon \)-independent estimates are obtained by taking \( u - \bar{u} \) as a test function where \( \bar{u}(t, \cdot) \) is a prolongation of \( \bar{u} \) in \( \Omega \setminus B(t) \) fulfilling the same boundary conditions as \( u \) and the constraint \( \text{div} \, \bar{u} = 0 \). To prove the estimates, we need time-independent estimates on this prolongation. These are obtained thanks to standard time-independent trace and lifting theorem applied to \( \bar{u}(t, \Psi^{-1}(t, x)) \) which is well defined in the non-moving non-deformable domain \( B(t) \) thanks to Hypothesis H2. We recover time-independent bounds thanks to the bounds we have on \( \Psi \). ■

Both results prove that the function \( G \) is well-defined from \( E \cap F \) to itself with
\[
E = L^\infty(0, T; H^1(\Omega)) \cap L^2(0, T; H^2(\Omega)) \quad \text{and} \quad F = L^\infty(0, T; \mathbb{H}^1(\Omega)) \cap L^2(0, T; \mathbb{H}^2(\Omega)).
\]
Moreover, when \( \alpha \) and \( \alpha' \) are solutions of (5.9) with the respective velocities \( u \) and \( u' \), and \( \mu \) and \( \mu' \) are solutions of (5.10) respectively with the viscosities \( \mu \) and \( \mu' \), and \( \mu = \Phi(\alpha) \), \( \mu' = \Phi(\alpha') \), we obtain
\[
\|(\alpha - \alpha')(t, \cdot)\|_{H^1(\Omega)}^2 \leq C \|u - u'\|_{L^2(0, T; H^1(\Omega))}^2,
\]
\[
\|(u - u')(t, \cdot)\|_{H^1(\Omega)} \leq C \|u'(t, \cdot)\|_{H^2(\Omega)} \|\mu - \mu'(t, \cdot)\|_{H^1(\Omega)},
\]
\[
\|(\mu - \mu')(t, \cdot)\|_{H^1(\Omega)} \leq C (1 + \|\alpha'(t, \cdot)\|_{L^\infty(0, T; H^1(\Omega))}) \|(\alpha - \alpha')(t, \cdot)\|_{H^1(\Omega)}.
\]
This leads to
\[
\|G(\alpha) - G(\alpha')(t, \cdot)\|_{H^1(\Omega)} \leq C \|u - u'\|_{L^2(0, T; H^1(\Omega))}
\]
\[
\leq C \left( \int_0^T \|u'(t, \cdot)\|_{H^2(\Omega)}^2 \|(\mu - \mu')(t, \cdot)\|_{H^1(\Omega)}^2 \, ds \right)^{1/2}
\]
\[
\leq C \|u'\|_{L^2(0, T; H^2(\Omega))} \|\mu - \mu'\|_{L^\infty(0, T; H^1(\Omega))}
\]
\[
\leq C \|u'\|_{L^2(0, T; H^2(\Omega))} \|\alpha - \alpha'\|_{L^\infty(0, T; H^1(\Omega))},
\]
where the constant \( C \) only depends on \( T, \Omega, f, \bar{u}, \alpha_0, \epsilon \) and \( \Phi \|_{L^\infty(\mathbb{R})} \).
Since
\[ \| \nabla \mu(t, \cdot) \|_{L^\infty(\Omega)} \leq C \| \Phi' \|_{L^\infty(\mathbb{R})} \| \alpha(t, \cdot) \|_{H^1(\Omega)}^{1/4} \| \alpha(t, \cdot) \|_{H^1(\Omega)}^{3/4}, \]
we obtain that \( \mu' \in L^{8/3}(0, T; W^{1, \infty}(\Omega)) \) and then that \( u' \in L^{8/3}(0, T; H^2(\Omega)) \). Finally,
\[ \| u' \|_{L^2(0, T; H^2(\Omega))} \leq t^{1/8} \| u' \|_{L^{8/3}(0, T; \mathbb{H}^2(\Omega))}. \]

There exists then a time \( T^* \in [0, T] \) such that
\[ \| (G(\alpha) - G(\alpha'))(t, \cdot) \|_{L^\infty(0, T^*; \mathbb{H}^1(\Omega))} \leq \frac{1}{2} \| \alpha - \alpha' \|_{L^\infty(0, T^*; \mathbb{H}^1(\Omega))}. \]

Moreover, the time \( T^* \) only depends on \( T, \Omega, f, \bar{u}, \alpha_0, \varepsilon \) and \( \| \Phi \|_{L^\infty(\mathbb{R})} \). Thanks to the estimates in Theorem 3 and to the estimates in Theorem 4 that are independent of \( \alpha \), we obtain the existence of a bound on \( \alpha \) valid for all \( t \in [0, T] \) and depending only on the data of the problem. Replacing the bounds on \( \alpha \) by this new one, we obtain a smaller time \( T^* \) valid starting from all \( t \in [0, T] \).

Since the function \( G \) is a contraction on \([0, T^*]\), we obtain that the recurring sequence \( (\alpha^n, u^n, p^n) \) converges to a weak solution on \([0, T^*]\), then on \([T^*, 2T^*]\), and finally on \([0, T]\). Moreover, since \( G \) is a contraction and thanks to the estimates we have, we prove the uniqueness of the solution.

### 5.2.1.2 Sketch of the Proof of Theorem 2

The solutions of (5.4)–(5.5) are obtained in the limit as \( \varepsilon \) tends to zero of the solution of (5.6)–(5.7). According to Theorem 1, we have that
- \( \alpha^\varepsilon \) is bounded independently from \( \varepsilon \) in the spaces \( L^\infty(0, T; \mathbb{H}^2(\Omega)) \), \( L^2(0, T; \mathbb{H}^3(\Omega)) \) and \( \mathbb{H}^1(0, T; H^1(\Omega)) \), which also implies that \( \alpha^\varepsilon \) and \( \nabla \alpha^\varepsilon \) are bounded in \( \mathbb{H}^1((0, T) \times \Omega) \),
- \( u^\varepsilon \) is bounded in \( L^q(0, T; V(\Omega)) \) for all \( 2 \leq q \leq +\infty \),
- \( \varepsilon^{-1/2} \| u^\varepsilon - \bar{u} \|_{L^2(0, T; B(\cdot))} \) is bounded for all \( 2 \leq q \leq +\infty \).

The \( \varepsilon \)-independent bounds makes it possible to extract subsequences still denoted \( (u^\varepsilon, p^\varepsilon, \alpha^\varepsilon) \) that converge weakly or weakly-* depending on the bounds and such that \( u^\varepsilon \) converges strongly to \( \bar{u} \) in \( \mathbb{H}^q(0, T; \mathbb{L}^2(B(\cdot))) \) for all \( 2 \leq q \leq +\infty \).

One of the keys to passing in the limit for non-linear terms of the equations consists in getting some strong convergence. This is obtained thanks to the bound on \( \alpha^\varepsilon \) and \( \nabla \alpha^\varepsilon \) in \( \mathbb{H}^1(O) \) which, thanks to Sobolev injection theorem in \( \mathbb{R}^d \), the strong convergence of a subsequence of \( \alpha^\varepsilon \) in \( W^{1,q}(O) \) with \( 2 \leq q \leq 4 \). We can now take the limit for \( u \) in \( B(\cdot) \) and use Equation (5.8) in the domain \( \Omega \setminus B(t) \) as well as for \( \alpha^\varepsilon \) in (5.9). Finally, the uniqueness of the solution can be gained by adapting the estimates that were used to prove that the function \( G \) defined in Section 5.2.1.1 is a contraction.
5.2.2 Numerical Analysis

This section presents the numerical algorithms specially developed to compute this mucus flow propelled by the ciliated epithelium cells. The equation set is rewritten as

\[
\begin{align*}
\frac{\partial \alpha}{\partial t} + u \cdot \nabla \alpha - \eta \Delta \alpha &= 0 \quad \text{in } \Omega, \\
\mu &= \Phi(\alpha) \quad \text{in } \Omega, \\
-\text{div}(2\mu D(u)) + \frac{1}{\varepsilon} B(t)(u - \bar{u}) &= f - \nabla p \quad \text{in } \Omega, \\
\text{div } u &= 0 \quad \text{in } \Omega,
\end{align*}
\]

(5.11)

where all the quantities have the same meaning as in the previous section. The $\varepsilon$ indices are removed in this part to improve the algorithm readability.

The first part of this section brings details on the computation of the cilia motion. The second part is dedicated to the computation of the mucins’ ratio $\alpha$ (first equation of (5.11)). The last part details the computation of the fluid velocity, namely the Stokes problem (last two equations of (5.11)).

5.2.2.1 Computation of the Cilia Beating

The cilia motion is decomposed into two phases: the recovery stroke and the effective stroke. In the recovery stroke, which lasts about 2/3 of the beating period, cilia are bending and moving backward, close to the bronchial wall and deep in the periciliary fluid layer. During the effective stroke the cilia are more straight and beat forward, penetrating in the mucus layer.

In order to numerically represent a coherent beating, each cilium is modeled by a filament of 0.3 µm diameter. The motion of this filament is governed by a transport equation set on a parametric curve which describes the filament center-line. Eqs. (5.12) are computed to obtain this, which is presented in Fig. 5.5, compared with the state-of-the-art experimental and numerical works.

Eventually, the beating remains planar in this study, and a small decay can be added between two adjacent cilia to create a metachronal synchronization:

\[
\frac{\partial L}{\partial t} + v(t) \frac{\partial L}{\partial \theta} = 0, \quad P(0, t) = 0, \\
L(0, t) = \partial_\theta P(0, t) = g(t) = (2\cos(2\pi t/T), 0, 1)
\]

(5.12)

where $\theta$ is the curve parameter, $P(\theta, t)$ is the curve position at time $t$, $L = \partial_\theta P$ and $v(t) = (1 + 8\cos^2(\pi(t + 0.25T)/T))/T$, with $T$ being the beating period. To obtain a metachronal synchronization, the same shift is added to both $g$ and $v$.

Eq. (5.12) enables the computation of the filament position at any time $t$. Then the characteristic function of $B(t)$, the volume occupied by the cilia array, can be deduced from the parametric curves, as well as the cilia velocity field $\bar{u}(t)$. These quantities are used in the following sections.

In all the computations, the counterforce applied by the fluid on the solid is not taken into account in the beating model. To our knowledge, this kind of computation and corresponding beating adaptation was only studied experimentally (Hill et al., 2010), and coherent modeling remains an open problem.
5.2.2.2 Numerical Computation of the Mucins Convection–Diffusion

The first two equations of (5.11) can be rewritten as an autonomous dynamical system. Indeed, the velocity can be represented a function of the mucins’ ratio through the viscosity, $u = F(\alpha)$:

$$\begin{align*}
\partial_t \alpha + F(\alpha) \cdot \nabla \alpha - \eta \Delta \alpha &= 0 \quad \text{in } \Omega, \\
\mu &= \Phi(\alpha) \quad \text{in } \Omega.
\end{align*}$$

(5.13)

The computation of this non-linear convection–diffusion equation can be difficult in a straightforward manner (finite difference discretization, for example) since two CFL stability conditions arise, one for each phenomenon (convection and transport). Hence the time step must usually be very small, imposing a large number of iterations and a large number of linear system assembling and resolution, which is computationally inefficient in 3D.

The viscous splitting (see Beale and Majda, 1981) of Eq. (5.13) enables the separation of the resolution of both operators $\partial_t + F(\alpha) \cdot \nabla$ and $\partial_t - \eta \Delta$, and allows the independent solution of both problems representing each phenomenon: a linear heat equation for diffusion (which is quite common to solve, particularly when $\eta$ is small enough to allow explicit computations, which is the case in mucus framework) and a non-linear convection equation. The splitting introduced in Chatelin et al. (2015) guarantees a second-order accuracy and permits alternat-
ing the resolution of diffusion and convection in order to benefit of diffusion regularizing effects after each sub-step of convection.

In order to compute the convection, a Lagrangian method is used (Cottet and Koumoutsakos, 2000), the mucin’s ratio is discretized on a particles cloud:

\[
\alpha(t) = \sum_{p=1}^{P} \alpha_p(t) \delta_{\xi_p(t)} v_p(t)
\]  

(5.14)

where \( \xi_p \) and \( v_p \) are respectively the position and the volume of particle \( p \) which carries a mucins’ ratio \( \alpha_p \). Using such a discretization transforms the convection PDE into a set of ODEs:

\[
\frac{d\xi_p}{dt} = u(\xi_p(t), t), \quad \frac{d\alpha_p}{dt} = 0, \quad \frac{dv_p}{dt} = 0.
\]  

(5.15)

In the last equation, particles’ volume does not change since the velocity field is incompressible.

The advantage of solving (5.15) instead of a convection PDE is that there is no CFL restriction, so large time steps can be achieved. Moreover, in (5.15), only the first equation needs to be computed, which is very computationally efficient and is well suited for 3D computations.

An important feature of the present Lagrangian method is that the velocity field is computed on a Cartesian grid, where the viscosity (and thus mucins’ ratio) needs to be known. Hence it is necessary to interpolate these quantities between the grid and the particles. To do that, a fourth-order interpolation kernel \( M_4' \) (Monaghan, 1985) is used. It guarantees fast computations since the computational cost is also linearly proportional to the number of particles (which is identical to the number of grid points) and accurate results due to the kernel’s third order and regularity.

The operator splitting combined with the Lagrangian computation of convection and accurate interpolations with well chosen kernels enables linear computations with respect to the number of grid points. In order to keep the feature, the computations of velocity also needs to be efficient.

5.2.2.3 An Iterative Projection for the Incompressible Stokes Problem

The computation of the Stokes problem at any given time \( t \) is considered now. The cilia position and velocity are already computed (so \( \bar{u}(t) \) and \( 1_B(t) \) are known) and the convection–diffusion of \( \alpha \) was achieved so \( \mu(t) \) is also computed. The last two equations of the set in (5.11) remain to be solved:

\[
\begin{align*}
-\text{div}(2\mu D(u)) + \frac{1_B(t)}{\varepsilon} (u - \bar{u}) &= f - \nabla p \quad \text{in } \Omega, \\
\text{div } u &= 0 \quad \text{in } \Omega \setminus B(t),
\end{align*}
\]  

(5.16)

To compute this problem an iterative projection method has been developed (Chatelin and Poncet, 2013). This class of methods has been widely used for non-stationary Navier–Stokes equations (Guermond et al., 2006). The principle of all these methods is quite similar: the first step consists in solving the momentum equation without the pressure gradient. Hence an
intermediate velocity field $u^*$ is introduced that is not required to satisfy the incompressibility condition, $\text{div } u = 0$.

A projection is then computed and the intermediate velocity is corrected in order to obtain a divergence-free velocity. This is performed by means of the projector $P(u^*) = u^* - \nabla \zeta$ where $\zeta$ is solution of $-\Delta \zeta = -\text{div } u^*$ in the whole domain, and satisfying the boundary condition $\partial \zeta / \partial n = u \cdot n$.

The main drawback of these methods is that the projector is usually computed by solving a Poisson problem with an homogeneous Neumann boundary condition. After the correction, the velocity field usually does not satisfy the original boundary condition (Guermond et al., 2006).

That the boundary conditions are not satisfied becomes more pronounced at low Reynolds numbers. This is the reason that projection methods developed for Navier–Stokes flows cannot be applied to the Stokes equations (Chatelin, 2013). To overcome this problem, a new iterative projection method was introduced in Chatelin and Poncet (2013). The algorithm is presented as follows:

1. (Initialization) $u^*_0 = u_0$ is provided.
2. (Fixed point iterations, where $i$ denotes the iteration number)
   (a) $u^*_i$ is known, as well as $u_i = P(u^*_i)$ and $\nabla \zeta_i = u^*_i - P(u^*_i)$,
   (b) (Prediction) $u^*_{i+1}$ is the solution to
   \[
   \begin{cases}
   -\mu \Delta u^*_{i+1} + \frac{1}{\rho} u^*_i - \nabla \zeta_i - \vec{u}(t) = f + (2D(u_i) + \text{div } u^*_i) \nabla \mu & \text{in } \Omega, \\
   u^*_{i+1} = g - \nabla \zeta_i & \text{on } \partial \Omega,
   \end{cases}
   \tag{5.17}
   
   \text{where the last term of (5.17) comes from the substitution } u^* = u + \nabla \zeta.$
   (c) (Correction) Compute $\zeta_{i+1}$, the solution to a Poisson problem
   \[
   \begin{cases}
   -\Delta \zeta_{i+1} = -\text{div } u^*_{i+1} & \text{in } \Omega, \\
   \partial \zeta_{i+1} / \partial n = 0 & \text{on } \partial \Omega,
   \end{cases}
   \tag{5.18}
   
   \text{and project, namely let } u^*_{i+1} = P(u^*_{i+1}) = u^*_{i+1} - \nabla \zeta_{i+1}.
   
   (d) Repeat steps 2a through 2c and increment $i$ until convergence is satisfied.
3. The solution to the Penalized Stokes problem is $u_{i+1}$.

This algorithm only involves the computation of elliptic problems. In particular, the computation of the projector can be achieved using an FFT or multigrid solver (Swarztrauber and Sweet, 1975; Adams, 1989) which are very powerful. Indeed, using a regular Cartesian grid, the structure of the discretized operator is known a priori and no matrix assembling or linear system resolution is required. Hence the computational cost is quasi-linear with respect to $N$, the number of computational grid points, that is to say, with a complexity $O(N \log N)$. Furthermore, it guarantees an accurate computation of boundary conditions, immersed solid velocity, inviscid velocity and non-homogeneous effects.

In order to conserve this computational efficiency, which is very attractive for 3D computations, the penalized Stokes equation (5.17) and the projection using FFT need to be computed with a similar computational cost. The next section describes a numerical method for the penalized Stokes equation using only FFTs.
5.2 NEWTONIAN MODELS

5.2.2.4 A Sherman–Morrison–Woodbury Approach to Compute Penalized Poisson Problems

To compute the Penalized Poisson problem of the first step of the previous iterative projection, a special algorithm has been developed (Chatelin and Poncet, 2014), based on the algebraic perturbation relation, the Sherman–Morrison–Woodbury formula (Hager, 1989). This formula enables the computation of the inverse of an invertible perturbed matrix

\[
(A + E^T CE)^{-1} = A^{-1} - A^{-1} E^T \left(C^{-1} + EA^{-1}E^T\right)^{-1} EA^{-1}.
\]  

(5.19)

The problem to solve (first step of the previous iterative projection) can be rewritten as follows (gathering all the terms depending on previous iteration \(k\) in the RHS):

\[
\begin{align*}
-\Delta u + cu &= f \quad \text{in } \Omega, \\
u &= 0 \quad \text{on } \partial\Omega,
\end{align*}
\]

(5.20)

where \(c\) represents the Penalization term and iteration subscripts were removed. Hence this problem can be seen as a Poisson problem perturbed in the cilia volume. Then, applying a generic discretization to (5.20), it can be exactly rewritten as an algebraic perturbation problem, and the SMW formula can be applied:

\[
(A + E^T CE)U = F \iff U = A^{-1}F - A^{-1}E^T \left(C^{-1} + EA^{-1}E^T\right)^{-1} EA^{-1}F.
\]  

(5.21)

In this equation, each product involving \(A^{-1}\) can be achieved without matrix assembling and inversion by using the Poisson solver. The key point is to invert the term inside parenthesis without matrix inversion and assembling, which is achieved with an iterative GMRES method. The global algorithm reads in four steps:

1. Solve the Poisson problem \(AW = F\).
2. Solve \(QZ = EW\) with \(Q = C^{-1} + EA^{-1}E^T\). This is performed by means of the GMRES method that requires the product evaluation \(Z \rightarrow QZ\) as follows:
   (a) Solve the Poisson problem \(A\Theta = E^T Z\);
   (b) Set \(QZ = C^{-1}Z + E\Theta\).
3. Solve the last Poisson problem \(AY = E^T Z\).
4. Update the solution \(U = W - Y\).

Steps 2a and 2b are computed at each Krylov iteration so the total number of Poisson problems to solve is \(2 + N_{Kry}\) where \(N_{Kry}\) is the typical size of the Krylov space. The other steps are negligible from a computational point of view: as \(C\) is a diagonal matrix, its inverse is immediately computed, restriction and inclusion matrices \(E\) and \(E^T\) are deduced from \(1_{B(t)}\) and never assembled.

The overall algorithm, presented in the previous sections has the following convergence features:

- A \(O(\sqrt{\varepsilon})\) convergence order for the Penalization method, which is a standard result (Carbou and Fabrie, 2003).
5. MODELING CYSTIC FIBROSIS AND MUCOCILIARY CLEARANCE

FIGURE 5.6 Snapshots of the mucus propulsion by a carpet of 25 ciliated epithelium cells at five different instants. The colormap shows the magnitude of the first component of the velocity field. At the beginning of the computation the cross-linked mucins ratio $\alpha$ is initialized according to Eq. (5.3). The periciliary fluid viscosity is equal to that of water ($10^{-3}$ Pa s) and the viscosity of the mucus is 0.1 Pa s. An isosurface of $\alpha = 0.5$ is presented in blue. In white, a passive particle clearly shows that the mucus is transported toward trachea. Panels (A) to (E) are snapshots at times $t = 3.33 \times 10^{-2}$, 0.77, 0.28, 0.29 and 0.88 s.

- A second-order temporal accuracy, obtained with the splitting algorithm (Chatelin et al., 2015).
- A second-order spatial accuracy which is given by the fast FFT solver (Swarztrauber and Sweet, 1975).
- A very negligible mass diffusion in particle transport and interpolation processes (Chatelin and Poncet, 2013).

5.2.3 Numerical Computations of the Mucus Propelled by Ciliated Epithelium

The first simulation presented involves a simplified epithelium cell composed of 25 cilia regularly arranged as a $5 \times 5$ array. Snapshots are presented in Fig. 5.6. This simulation was
5.2 NEWTONIAN MODELS

TABLE 5.1  Reference parameters for the numerical simulations
(gathered from Fahy and Dickey, 2010; Sanderson and Sleigh, 1981; Smith et al., 2008; Thiriet, 2012)

<table>
<thead>
<tr>
<th>Name</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cilia length</td>
<td>8 µm</td>
</tr>
<tr>
<td>Cilia diameter</td>
<td>0.3 µm</td>
</tr>
<tr>
<td>Periciliary fluid viscosity ($\eta_{PCF}$)</td>
<td>0.001 Pa s</td>
</tr>
<tr>
<td>Viscosity ratio ($\beta = \eta_{ML}/\eta_{PCF}$)</td>
<td>100</td>
</tr>
<tr>
<td>Viscosity transition stiffness ($\gamma$)</td>
<td>10</td>
</tr>
<tr>
<td>Transition length ($\delta$)</td>
<td>7 µm</td>
</tr>
<tr>
<td>Airway surface liquid height ($H_z$)</td>
<td>13 µm</td>
</tr>
<tr>
<td>Beating frequency</td>
<td>10 Hz</td>
</tr>
<tr>
<td>Mucins diffusion ($\nu$)</td>
<td>$10^{-11}$ m$^2$s$^{-1}$</td>
</tr>
</tbody>
</table>

performed with the parameters gathered of Table 5.1. This figure presents an isosurface of cross-linked mucins ratio, velocity magnitude in the proximal direction and a passive black particle, advected by the flow. On average, this passive tracer travels forward (in the proximal direction to the trachea) which shows that the mucociliary clearance is effective.

The second simulation (see Fig. 5.7) presents the mucociliary transport with a carpet of 300 cilia, still using the parameters of Table 5.1. This computation presents the effect of the metachronal synchronization (when a shift is added in the beating between adjacent cilia, creating the metachronal wave) for a large number of cilia. This figure shows that the metachronal wave travels backward, generating a positive first velocity component wave in the mucus layer whereas the velocity is zero or lightly negative in the periciliary fluid layer.

As in the previous figure, the passive white particle clearly shows that the upper part of the airway surface liquid travels forward, thus the mucociliary clearance is efficient. This simulation shows that a “leading cavity” effect is created when cilia are in recovery stroke, maximizing the mucociliary transport. These results, also reported in Hussong et al. (2011) using simplified simulations, help to validate the present methodology.

5.2.4 Phenomena Analysis

In order to quantify the mucociliary clearance efficiency, the mean velocity of the mucus layer is averaged over several beating periods $T$:

$$U = \frac{1}{3TH_xH_y(H_z - \delta)} \int_0^{3T} \int_0^{H_z} \int_0^{H_y} \int_0^{H_x} u_x(x, y, z, t)dx dy dz dt$$

(5.22)

where $\delta$ is the transition height between PCL and mucus, introduced in Eq. (5.3).

Using different parameter sets, this quantity enables the comparison between different simulations (Chatelin and Poncet, 2016). This can be used in the identification of the meaningful parameters and can help to identify the influence of pathological situations in mucociliary transport. In this study, the following meaningful parameters were identified:

- The beating frequency $2\pi / T$ is an important parameter of the modeling. Indeed, as a consequence of the time-dilatation of the quasi-static Stokes equations, the mucociliary transport
FIGURE 5.7 Snapshots of the mucus propulsion by a carpet of 300 ciliated epithelium cells at five different times. Colormap and fluid features are the same as in Fig. 5.6, except that the isosurface level is $\alpha = 0.65$. Panels (A) to (E) are snapshots at times $t = 6.67 \times 10^{-3}$, 0.2, 0.23, 0.35 and 0.5 s.

It was also demonstrated that the cilia length is very important. In fact, when cilia are too small the mucociliary clearance is deteriorated. This paper shows that it can be divided by 10 if cilia are 3 $\mu$m long instead of 9. It was demonstrated that the most efficient result is
obtained when cilia penetrate the mucus layer during the effective stroke and remain in
the periciliary fluid layer during the recovery stroke. This result confirms this assumption
formulated in Liron and Rozenson (1983) and contradicts the results of Fulford and Blake
(1986).

• As expected, the viscosity ratio between mucus and periciliary fluid has also a great impor-
tance: when the mucus becomes highly viscous, the mucociliary clearance is also deterio-
rated, it can be divided by two when the mucus is 1000 times more viscous than periciliary
fluid layer. It is important to notice that the most efficient mucociliary clearance is not
obtained for a ratio of 1 (meaning an airway surface liquid only composed of periciliary
fluid) but when the mucus is about 10 times more viscous. It means that the mucociliary
clearance permits to handle the accumulation of cross-linked mucins in the mucus layer,
ensuring a good protection of the airways.

• The transition length $\delta$ between mucus and periciliary fluid layer is also very important. It
is the equilibrium between both these characteristic lengths which leads to the penetration
(or not) of the cilia tips in the mucus during the effective stroke. In particular, when this
transition length is too small (this is representative of the case in which nearly all of the
airway surface liquid is composed of mucus) the mucociliary clearance can be divided
by 2.5.

• Finally, the transition stiffness $\gamma$ between the periciliary fluid layer and the mucus is also
an important parameter. It means that to achieve an efficient mucociliary clearance, two
layers have to be clearly identified. This also enables the penetration in the mucus only
during the effective stroke.

5.3 RHEOLOGY OF MUCUS AND NON-NEWTONIAN MODELS

In this section, the methods and data available in the literature are presented, followed by
our own methods and results. The power law and Carreau law are then considered the
best fitting models, and introduced in the mathematical model and in the numerical scheme.
A discussion confronting experimental data and results from numerical simulation is finally
performed, with a description of the medical outcomes.

5.3.1 Rheometry Data on Lung Mucus from the Literature

Looking at studies concerned with rheological characterizations of pulmonary mucus, two
points are noticeable. First, two kinds of equipment are used: as expected, classical rheome-
ters, but also magnetic micro-rheometers. Second, rheological characterizations are often
limited to the determination of the dynamic storage modulus ($G'$) and loss modulus ($G''$)
for two frequencies, 1 and 100 rad/s, which are supposed to be characteristic frequencies of
mucociliary clearance and cough clearance, respectively (App et al., 1998).

5.3.1.1 The Different Measurement Methods

Considering rheological measurements on lung mucus, there are almost as many studies with
magnetic micro-rheometers as with classical rheometers. The magnetic micro-rheometer uses
either a single steel bead (App et al., 1998; Redding, 2008) or multiple magnetite particles (Edwards and Yeates, 1992; Ziemann et al., 1994) positioned in a few µL of fluid and oscillated by means of an electromagnetic field to obtain a sinusoidal force. In the first case, the bead diameter ranges from 50 to 100 µm while particles dimension ranges from 2 to 6 µm in the second case. The motion of the single bead or of the particles embedded in the fluid is then quantified, generally by fast image processing techniques; see King and Macklem (1977) and Ziemann et al. (1994) for the detail of rheological parameters extraction. Note that King and Macklem (1977) have incorporated Thomas and Walters (1966) results to take into account elasticity of the fluid. This technique has largely been used for respiratory mucus (Jeanneret-Grosjean et al., 1988; Rubin et al., 1990; App et al., 1998; Zayas et al., 1999; Redding, 2008).

The main advantage of the micro-rheometer lies in the very small volume of fluid that is necessary (a few µL). This is of particular interest for mucus that is generally not available in large quantities except for major pathologies such as cystic fibrosis. It is important to note that in general, mucus type fluids are not homogeneous, and that larger sample volumes would allow a more representative measurement of the global rheological behavior and a better precision. Additional drawbacks are that only oscillatory experiments are possible with such a micro-rheometer and that the flow is not a simple shear flow as it is theoretically required for rheometry (Barnes et al., 1989).

Classical rheometry has predominantly been used in recent years (Sanders et al., 2000; Dawson et al., 2003; Nielsen et al., 2004; Innes et al., 2009; Serisier et al., 2009; Daviskas et al., 2010; Tomaiuolo et al., 2014). It is performed using controlled-stress rheometer equipped with cone and plate geometry or parallel plates geometry. These are generally chosen to minimize the fluid volume: small diameter/small angle for the cone (1° or 2°), or small gap for parallel plates (between 50 and 250 µm when specified).

Note that the term “micro-rheology” may be confusing as it is also used to represent length scales below 0.1–0.3 µm, for instance, to examine virus or nano-particle motion through mucus (Sanders et al., 2000; Dawson et al., 2003; Lai et al., 2009; Schuster et al., 2013). Viscosity experienced by these objects is close to that of water: it is linked to the mesh size of the network structure of mucus which is larger than (or similar to) the object size. Nevertheless, for the mucus clearance process larger length scales are relevant. The viscosity has to be considered macroscopic, which is anticipated to greatly exceed the viscosity of water and is associated to a complex rheological behavior.

5.3.1.2 Mucus Rheology from Literature

A general presentation of mucus, its role, chemical structure and physiological properties can be found in Cone (2005). Reviews of the rheological properties are presented by Yeates et al. (1997) and Picout and Ross-Murphy (2003). Considering human airway mucus, few data are available in the literature for steady state viscosity while data for dynamic viscosity \( \eta' = G'/\omega \) or complex viscosity \( |\eta^*| = |G^*|/\omega \), from oscillatory measurements, are more numerous. When it is reported, the measurement temperature is 20, 25, or 37°C. This might appear as a large discrepancy but variances caused by temperature differences are small compared to those from the non-Newtonian feature, as it was demonstrated by Tomaiuolo et al. (2014).

When a rheogram is proposed, all authors agree to recognize a strong shear thinning behavior. Some authors (Moores et al., 1992; Dawson et al., 2003; Suk et al., 2009) observed
5.3 RHEOLOGY OF MUCUS AND NON-NEWTONIAN MODELS

a tendency to a Newtonian plateau at low shear rates with a transition towards power-law behavior between 0.1 s⁻¹ and 1 s⁻¹, depending on authors. On the other hand Yeates et al. (1997) did not observe any trend to a plateau although exploring shear rate from 0.2 s⁻¹ upwards. The same observation was made by Puchelle et al. (1985, 1987) for shear rate starting from 0.01 and 0.1 s⁻¹, respectively. In the shear thinning region, the behavior can be modeled by a power-law \( \tau = k \dot{\gamma}^n \), where \( \tau \) and \( \dot{\gamma} \) are the shear stress and the shear rate respectively, \( k \) the fluid consistency and \( n \) the power-law index. Values for \( k \) and \( n \) given in literature, or extracted from literature data, are reported in Table 5.2. Note the low values of the power-law index (between 0.04 and 0.26) which are characteristic of a strong shear-thinning behavior.

Thixotropy is the property of a fluid to show a reversible decreasing viscosity when submitted to a constant shear rate (i.e. observation of a decreasing shear stress as a function of time). Note that thixotropy is sometimes confused with shear-thinning, although rheological definition is unambiguous (Picout and Ross-Murphy, 2003). Thixotropy leads to a hysteresis phenomena that has been observed on viscosity measurements at increasing shear rates followed by measurements at decreasing shear rates (Nielsen et al., 2004; Tomaiuolo et al., 2014). Conversely, note that hysteresis is not necessarily evidence of thixotropy because elasticity may also be responsible for hysteresis, especially when the characteristic time of the fluid is comparable to (or larger than) the measurement time scale. References to a characteristic time for mucus are not numerous in the literature: Besseris and Yeates (2007) gave 42 s for canine tracheal mucus, while for human mucus Moores et al. (1992) obtained 20 s from creep tests, and Nielsen et al. (2004) estimated it to be of the order of thousands of seconds from oscillatory measurements. Academic thixotropy measurements were realized by Puchelle et al. (1985, 1987) who observed a shear stress overshoot after a shear rate step. Such an overshoot is linked to elasticity, but successive experiments with decreasing rest time after previous shearing show a decreasing overshoot. This has allowed Puchelle et al. to identify the presence of thixotropy for reconstituted mucus (Puchelle et al., 1985) and native mucus (Puchelle et al., 1987).

A few authors have identified a viscoplastic behavior with a yield stress varying between 0.05 to 0.26 Pa (Edwards and Yeates, 1992) until 100 Pa (Silberberg, 1983). For instance, this last value was used by Craster and Matar (2000) to numerically analyze the surfactant transport of mucus film in the pulmonary airways. More recently, Broughton-Head et al. (2007) proposed a value of 14 Pa for cystic fibrosis mucus. As far as we know, these are the only publications identifying a viscoplastic behavior and giving yield stress values. This can be connected to specific rheometers used or to the conduction of the rheological characterizations.

On the other hand, elasticity has been clearly identified in the literature. This is due in part to the widespread use of the magnetic micro-rheometer which is restricted to small-amplitude oscillatory shear experiments. The storage modulus \( G'(\omega) \) and the loss modulus \( G''(\omega) \) are the most common data encountered in the literature at the two frequencies \( \omega = 1 \) and \( \omega = 100 \) rad/s. The moduli \( G' \) and \( G'' \) are the imaginary and real parts of the complex modulus \( G^* = \tau/\gamma \), where \( \gamma \) is the deformation. A phase shift between shear stress and shear rate \( \dot{\gamma} \), i.e. a non-zero \( G' \) value, demonstrate the elastic feature of the fluid.

Data are expressed in terms of modulus or, in equivalent form, using the complex modulus \( G^* \) and the loss tangent \( \tan \delta = G''/G' \) (not to be confused with the PCL height in the previous sections), or \( G' \) and the so-called dynamic viscosity \( \eta' = G''/\omega \). The values of \( G' \) and \( G'' \) for two specific values of the frequency, 1 and 100 rad/s, are often reported in
### TABLE 5.2: Mucus rheological characterization, power-law parameters; CR, Classical rheometer; \( \mu R \), Magnetic \( \mu \)-rheometer; CF, Cystic Fibrosis; Br, Bronchiectasis; COPD, Chronic Obstructive Pulmonary Disease; CB, Chronic Bronchitis; LS, Light Smokers; HS, Heavy Smokers; XS, ExSmokers; C, Cancer; FEV1\%, ratio of the Forced Expiratory Volume in 1 s (FEV1) to the Forced Vital Capacity (FVC)

<table>
<thead>
<tr>
<th>Source</th>
<th>Alias</th>
<th>Rheometer</th>
<th>Pathology</th>
<th>Additional information</th>
<th>( n )</th>
<th>( k )</th>
<th>( n^* )</th>
<th>( k^* )</th>
</tr>
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<tbody>
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<td>CR</td>
<td>CR</td>
<td>Chronic airway hypersecretion</td>
<td></td>
<td>0.18</td>
<td>18.38</td>
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<td>HL6</td>
<td>( \mu R )</td>
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<td></td>
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<td></td>
<td>0.19</td>
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<td>( \mu R )</td>
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<td></td>
<td></td>
<td>–</td>
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<td>( \mu R )</td>
<td>Healthy After administration of Atropine</td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
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<td>Rubin et al. (1990)</td>
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<td>CF</td>
<td>Healthy Taken from endotracheal tube at the end of a surgery</td>
<td></td>
<td></td>
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<td></td>
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<td>18.62</td>
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<td></td>
<td></td>
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<td>6.92</td>
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<td>18.00</td>
<td>0.17</td>
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<td>CF</td>
<td>–</td>
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<td>Redding (2008)</td>
<td>M( \mu R )</td>
<td>Br</td>
<td>Idiopathic bronchiectasis (children)</td>
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<td></td>
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<td>CF</td>
<td>Children</td>
<td></td>
<td></td>
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<td>0.16</td>
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<td>Redding (2008)</td>
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<td>CB</td>
<td>Adults</td>
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<td>0.14</td>
<td>53.03</td>
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<td>Innes et al. (2009)</td>
<td>CR</td>
<td>CR</td>
<td>Healthy Sputum induction performed with nebulized 3% saline for 20 min</td>
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</tr>
<tr>
<td>Serisier et al. (2009)</td>
<td>CFL1</td>
<td>COPD</td>
<td>Clinically stable</td>
<td></td>
<td></td>
<td></td>
<td>0.34</td>
<td>1.29</td>
</tr>
<tr>
<td>Suk et al. (2009)</td>
<td>CR</td>
<td>CF</td>
<td>–</td>
<td></td>
<td>0.16</td>
<td>2.42</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Daviskas et al. (2010)</td>
<td>CR</td>
<td>Br</td>
<td>Clinically stable</td>
<td></td>
<td>0.41</td>
<td>36.31</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Daviskas et al. (2010)</td>
<td>CR</td>
<td>Br</td>
<td>Clinically stable</td>
<td></td>
<td>0.45</td>
<td>24.76</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Schuster et al. (2013)</td>
<td>CR</td>
<td>Healthy</td>
<td>Taken from endotracheal tube at the end of a surgery</td>
<td></td>
<td></td>
<td></td>
<td>0.16</td>
<td>12.97</td>
</tr>
<tr>
<td>Tomaiuolo et al. (2014) according to FEV1</td>
<td>CFL2</td>
<td>CF</td>
<td>FEV1% &gt; 50%</td>
<td></td>
<td>0.23</td>
<td>2.18</td>
<td>0.18</td>
<td>1.40</td>
</tr>
<tr>
<td>Tomaiuolo et al. (2014) according to FEV1</td>
<td>CFL3</td>
<td>CF</td>
<td>30% &lt; FEV1% &lt; 50%</td>
<td></td>
<td>0.21</td>
<td>12.81</td>
<td>0.17</td>
<td>2.25</td>
</tr>
<tr>
<td>Tomaiuolo et al. (2014) according to main colonization</td>
<td>CFL4</td>
<td>CF</td>
<td>FEV1% &lt; 30%</td>
<td></td>
<td>0.26</td>
<td>6.87</td>
<td>0.16</td>
<td>2.42</td>
</tr>
<tr>
<td>Tomaiuolo et al. (2014) according to main colonization</td>
<td>CR</td>
<td>CF</td>
<td>S. aureus</td>
<td></td>
<td>0.20</td>
<td>3.04</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Tomaiuolo et al. (2014) according to main colonization</td>
<td>CR</td>
<td>CF</td>
<td>P. aeruginosa</td>
<td></td>
<td>0.17</td>
<td>14.10</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Tomaiuolo et al. (2014) according to main colonization</td>
<td>CR</td>
<td>CF</td>
<td>B. cepacia and S. maltophilia</td>
<td></td>
<td>0.14</td>
<td>10.86</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>
the literature because they were proposed by some authors (Jeanneret-Grosjean et al., 1988; Zayas et al., 1990, 1999; Shah et al., 1996; App et al., 1998; Innes et al., 2009) to define a mucociliary clearability index (conventionally called MCI) and a cough clearability index (CCI), which respectively quantify the clearability by normalized ciliary function (using values at 1 rad/s) and by cough (using values at 100 rad/s). As far as we know, the mucociliary clearability index was first proposed by Jeanneret-Grosjean et al. (1988). MCI and CCI are then mentioned in Zayas et al. (1990). Note that these indices were also used by Tomaiuolo et al. (2014) at a frequency of 10 rad/s for CCI. These indices are of particular interest in the evaluation and quantification of the influence of pathologies on the mucus clearance for smokers and lung cancer patients (Zayas et al., 1990, 1999), for asthma (Innes et al., 2009), and for the influence of treatments or physiotherapy (Shah et al., 1996; App et al., 1998).

In the study of the flow of mucus, the essential parameter is the viscosity. One can ask whether the Cox–Merz rule is sufficient to evaluate \( \eta(\dot{\gamma}) \) using oscillatory measurement data.

The Cox–Merz rule is considered as very useful when only linear viscoelastic data are available (Bird et al., 1977). It is expressed as:

\[
|\eta^*(\omega)| = \sqrt{\eta'(\omega)^2 + \eta''(\omega)^2} = \eta(\dot{\gamma})|_{\dot{\gamma}=\omega}. \tag{5.23}
\]

The Cox–Merz rule is well-adapted for polymer solutions with entangled network structure. It fails (with generally \(|\eta^*| > \eta\)) for cross-linked systems or for highly concentrated entangled systems. Cross-linked polymers are characterized by the non-existence of a Newtonian plateau for \( \eta(\dot{\gamma}) \) and a weak evolution of elastic and viscous modulus with frequency. Based on such observations, numerous authors think that mucus is of cross-linked type (Puchelle et al., 1985; Innes et al., 2009; Schuster et al., 2013) but Yeates et al. (1997) developed arguments for an entangled “sliplinked” strand system. Looking at Table 5.2, there are only six cases where the power-law parameters \((n, k)\) and \((n^*, k^*)\) (from power-law model applied to \( \eta^*(\omega) \) available data) can be estimated and compared. The consistency \( k \) is underestimated when assimilated to \( k^* \) for five of the six cases, while no real tendency can be observed on \((n, n^*)\) data (see Fig. 5.8). This point will be re-examined later.

Information related to pathology are presented in Fig. 5.9. For this representation, the rheological variables \( |G^*| \) and \( \tan\delta \) were chosen as they are often considered as relatively independent from one another while \( G' \) and \( G'' \) are not (Jeanneret-Grosjean et al., 1988; King, 1980). Nevertheless, \( |G^*| \) varies with respect to viscosity while \( \tan\delta \) compares viscous and elastic response of the fluid (with \( \tan\delta \) infinite for purely viscous fluids). Concerning healthy patients, it is noticeable that the two points corresponding to the lower \( |G^*| \)-value were obtained after nebulization with a saline solution to induce sputum. This probably led to the thinning of the mucus and also helps to explain these lower values as well as the large \( \tan\delta \) value.

These comparisons have to be considered cautiously because conditions differ between studies (how mucus is taken and prepared, how rheological characterizations are conducted, for example). It is noteworthy to point out that mucus samples of patients with pathology do not strongly differ from samples of healthy patients (except in some cases of bronchiectasis and COPD). On this graph, the majority of data corresponds to stable patients, or healthy, one case corresponds to exacerbated situation and no information is given for five cases. In a
FIGURE 5.8 Comparison between consistency $k$ and power-law index $n$ issued from viscosity curves and estimations ($k^*, n^*$) issued from the Cox–Merz rule. Note that each point from literature is a mean value while a point marked “this study” corresponds to one of the tested mucus. The solid line denotes the identity.

FIGURE 5.9 Rheological parameters $G^*$ and $\tan \delta$ (measured at 1 rad/s) for different pathologies. Symbols are the same as in Table 5.2. All cases with simple symbols concern studies of patients in a stable state. Symbols with a surrounding square indicate a study for patients in exacerbation or a study without information.

A study on rheological properties of CF mucus, King (1981) proposed to classify mucus from its visual aspect in three categories (linked to the nature and to the degree of infection): mucoid, muco-purulent and purulent. Comparing $G'$ and $\eta'$ for three frequencies, he observed that rheological properties for mucoid and muco-purulent human CF mucus are similar to control canine tracheal ones while purulent mucus showed higher values and a diminution of $\tan \delta$ (i.e. higher elasticity). This was also underlined by Deneuville et al. (1997), Lopez-Vidriero and Reid (1978). Although this distinction between mucus seemed no longer considered for several years, more recent studies have confirmed the interest of this observation, using stable/exacerbated distinction rather than visual criteria: Serisier et al. (2009) and Tomaiuolo et al. (2014) for CF sputum, and Tabatabaei et al. (2015) for COPD.
5.3.2 Mucus Sample Analysis and Rheological Results

5.3.2.1 Mucus Samples Collecting
Consecutive adult patients with cystic fibrosis (CF) bronchiectasis (Br) were recruited when admitted in the regional adult CF center of Toulouse. After information and oral consent, sputum samples were collected under supervision of a chest physiotherapist. Patients were classified as either having a stable status or having an exacerbation. The classification of exacerbation represented an increased cough or an increased bronchorrhea or purulence sputum, and/or an increased dyspnea, and/or a tiredness and/or a loss of weight). Sputum samples were frozen at −20°C before rheological analysis.

5.3.2.2 Mucus Sample Analysis
In this study rheological characterizations were evaluated on a Mars III rheometer (Thermo Sc.) equipped with a cone and plate geometry (1° angle, 35 mm diameter) which requires 0.2 mL of fluid. Measurements were conducted at 37°C. The lower plate Peltier system for temperature regulation was completed by a sample hood that was custom made in-house with a specific temperature regulation and wet sponges to saturate the measuring chamber and prevent evaporation of the sample. As previously mentioned, samples were stored frozen at −20°C. The influence of freezing on the rheological properties of mucus has previously been investigated by Rubin et al. (1990), Sanders et al. (2000), Gastaldi et al. (2000) and Serisier et al. (2009) for different conditions (temperature, freezing duration). These authors all conclude that freezing does not affect rheological behavior. This result was confirmed in the current study on three samples that were characterized when fresh and again at 21 or 31 days under frozen conditions. Mucus samples were thawed keeping them in a water–ice mixture just before and during measurements to prevent degradation (Sanders et al., 2000).

Once a sample was loaded in the rheometer, the following protocol was followed: an initial dynamical strain sweep at a fixed frequency of 1 Hz was conducted to determine the linear domain. Then the storage modulus \( G'(\omega) \) and the loss modulus \( G''(\omega) \) were determined using an oscillatory frequency sweep for a fixed strain in the linear domain (between 1 and 3% according to the sample). Then, the sample was submitted to very large deformations in which the viscosity \( \eta(\dot{\gamma}) \) was determined by a steady-shear flow test for shear rates in the range from \( 10^{-3} \) to \( 10^3 \) s\(^{-1}\). Careful attention was paid to reaching the steady state regime for cases with low shear rate values. Additional measurements were sometimes conducted to examine the stability of a sample. A power-law model \( \eta = k\dot{\gamma}^{n-1} \) \( (k, \text{consistency; } n, \text{power-law index}) \) was used to fit the rheological data. When the sampled volume was sufficient, two or three different tests were performed.

5.3.2.3 Rheological Results
Initially a few samples were used in order to determine the measurement strategy. Then 15 mucus samples were analyzed. Two of the samples were rejected because they were visually too heterogeneous and consequently gave rise to a large dispersion in viscosity and modulus values. The remaining 13 mucus samples were then considered for data analysis.

To examine thixotropy, experiments were conducted that were based on the observation of the shear stress overshoot after a shear rate step (similar to those of Puchelle et al., 1987). Because the results of those experiments were found to be inconclusive, another approach was
adopted. On one of the mucus samples (CFM7), viscosity measurements were conducted on four samples while on the fifth one, a large amplitude (300%, i.e. far from the linear domain) oscillatory sequence (LAOs) was first applied during 60 min. After that a viscosity measurement was done. The results are reported in Fig. 5.10A. The viscosities for samples S1, S2, S3 and S5 reveal the variability between samples and between measurements. Fig. 5.10 reports the data indicating the three measurement numbers. For measurements 2 and 3 (Meas. 2 and Meas. 3), only small amplitude oscillatory measurements were conducted before (strain sweep and/or frequency sweep). No specific classification linked to solicitations to which the sample was previously submitted appears in these results, especially after a strong and long solicitation such as 60 min of LAOs (S4, Meas. 2 and Meas. 3). Fig. 5.10B presents steady-state shear viscosity curves established for mucus CFM2, CFM6 and BM12. For each of them, two or three measurements were conducted, thus illustrating both the dispersion observed on the same mucus and the variability observed between mucus. The dispersion observed on all these series are similar, which implies that thixotropy is weak and comparable here to the natural dispersion arising from heterogeneity of samples and from the measurement difficulties.

As previously reported (Chatelin et al., submitted for publication), a strongly shear-thinning behavior is observed all over the shear rate range studied. Mucus samples were characterized on a wide shear rate range starting at $10^{-3}$ s$^{-1}$ and no trend to a Newtonian plateau was observed, corroborating the results of Puchelle et al. (1985, 1987) and Yeates et al. (1997). Note that the same observation can be made on the viscosity calculated from the Cox–Merz rule, i.e. from data for which the fluid has only been subjected to small deformations (in this case, the equivalent shear rate range covered is [0.01; 30] s$^{-1}$). This behavior is commonly associated with cross-linked polymers and is sometimes referred to as “yield stress” behavior (leading to the commonly used term “viscoelastic gel”). While data for shear rates as low as $10^{-3}$ s$^{-1}$ were conducted, the $\tau(\dot{\gamma})$ curves did not show any trend to a yield stress.
5.3 RHEOLOGY OF MUCUS AND NON-NEWTONIAN MODELS

### TABLE 5.3
Power-law parameters, relaxation time and power-law parameters from Cox–Merz rule from collected samples. \((n, k)\) data are from Chatelin et al. (submitted for publication)

<table>
<thead>
<tr>
<th>CFM1</th>
<th>CFM2</th>
<th>CFM3a</th>
<th>CFM3b</th>
<th>CFM4</th>
<th>CFM5</th>
<th>CFM6</th>
</tr>
</thead>
<tbody>
<tr>
<td>(n)</td>
<td>0.20</td>
<td>0.39</td>
<td>0.32</td>
<td>0.39</td>
<td>0.30</td>
<td>0.30</td>
</tr>
<tr>
<td>(k)</td>
<td>5.80</td>
<td>0.45</td>
<td>1.24</td>
<td>1.60</td>
<td>3.33</td>
<td>2.37</td>
</tr>
<tr>
<td>(T_c) (s)</td>
<td>(&gt;20)</td>
<td>(\gg20)</td>
<td>(\gg20)</td>
<td>(-)</td>
<td>(&gt;20)</td>
<td>(&gt;20)</td>
</tr>
<tr>
<td>(n^*)</td>
<td>0.17</td>
<td>0.07</td>
<td>0.25</td>
<td>0.04</td>
<td>0.15</td>
<td>0.23</td>
</tr>
<tr>
<td>(k^*)</td>
<td>24.6</td>
<td>3.08</td>
<td>0.98</td>
<td>8.45</td>
<td>6.32</td>
<td>9.92</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>CFM7</th>
<th>CFM8</th>
<th>CFM9</th>
<th>CFM10</th>
<th>CFM11</th>
<th>BM12</th>
</tr>
</thead>
<tbody>
<tr>
<td>(n)</td>
<td>0.21</td>
<td>0.23</td>
<td>0.37</td>
<td>0.36</td>
<td>0.22</td>
</tr>
<tr>
<td>(k)</td>
<td>3.10</td>
<td>3.53</td>
<td>0.51</td>
<td>2.17</td>
<td>4.75</td>
</tr>
<tr>
<td>(T_c) (s)</td>
<td>(\approx100)</td>
<td>(\approx100)</td>
<td>(\approx50)</td>
<td>(\approx30)</td>
<td>(&gt;20)</td>
</tr>
<tr>
<td>(n^*)</td>
<td>0.17</td>
<td>0.20</td>
<td>0.28</td>
<td>0.20</td>
<td>0.21</td>
</tr>
<tr>
<td>(k^*)</td>
<td>6.43</td>
<td>3.28</td>
<td>2.20</td>
<td>1.85</td>
<td>2.92</td>
</tr>
</tbody>
</table>

**FIGURE 5.11** Dynamic oscillatory shear tests (filled symbols, elastic modulus \(G'\); hollow symbols, viscous modulus \(G''\)): (A) strain amplitude sweep, (B) frequency sweep.

A power-law model was fitted on the shear rate range \([0.1; 100]\) s\(^{-1}\) which is representative of shear rate involved in the flow. The power-law parameter (index \(n\) and consistency \(k\)) are reported in Table 5.3 (from Chatelin et al., submitted for publication for \((n, k)\) values). The power-law indexes \(n\) are in the range 0.2 to 0.4 and are thus similar to values obtained by Puchelle et al. (1987), Nielsen et al. (2004), Suk et al. (2009) and Tomaiuolo et al. (2014) while Moores et al. (1992) and Dawson et al. (2003) observed lower \(n\)-values. Values of consistency \(k\), and then for viscosity, are also similar to the values published in the literature. The large shear rate range explored in this study gives an appropriate description of viscosity for the present study, which means this range is coherent with the shear rates observed in the numerical simulations.

Oscillatory measurements are presented in Fig. 5.11. Strain sweeps are presented for four mucus samples (Fig. 5.11A) and clearly show a large linear domain as it extends until 5 to 10%. Fig. 5.11B presents the storage modulus \(G'\) and the loss modulus \(G''\) from a frequency sweep. These curves clearly display the viscoelastic character of mucus: exploring the low fre-
quantities (until $10^{-3}$ Hz, i.e. more than 1000 s for a point), the $G'(\omega)$ and $G''(\omega)$ curves present the typical viscoelastic frequency-evolution: although slightly varying with frequency, these curves cross each other at a frequency $\omega_c$ which allows the determination of a characteristic times $T_c = 1/\omega_c$. This cross frequency was not reached for each mucus sample, but values or estimates are reported in Table 5.3. They range from 20 up to 100 s or more, thus highlighting the precautions to be taken for measurements that must correspond to a steady regime.

As a great number of studies only consider dynamic measurements, it is of interest to examine the validity of steady shear viscosity estimated from these measurements. For polymer solutions and polymer melts, the Cox–Merz rule (Eq. (5.23)) is considered as an acceptable way to evaluate viscosity. Parameters $k^*$ and $n^*$ of the power-laws fitted on $|\eta^*(\omega)|$ data are reported in Table 5.3 and presented in Figs. 5.8A–B for $k$ vs $k^*$ and $n$ vs $n^*$, respectively.

Results are slightly different from those extracted from the literature: for consistency, which can be regarded as viscosity at 1 s$^{-1}$, $k^*$ is similar or higher than $k$. This is coherent with what is known on the validity of the Cox–Merz rule which overestimates the shear viscosity (Picout and Ross-Murphy, 2003). Concerning the power-law index, $n^*$ is similar to $n$ for five cases but significantly lower for others, i.e. indicates a more shear-thinning behavior. Considering both $k^*$ and $n^*$, this signifies that the difference between dynamic viscosity and shear viscosity becomes increasingly large with decreasing shear rates: considering the viscosity at 1 s$^{-1}$, a relative differences of more than 100% is observed 7 times out of 13. For a shear rate of 0.01 s$^{-1}$, this score reaches 9 out of 13.

Nevertheless, a power-law index of 0.2 (resp. 0.4) leads to changes in viscosity in a ratio 6.3 (resp. 4) over an order of magnitude on shear rate. It follows that variations of viscosity in such a mucus flow are most of the time considerably larger than variations emanating from the discrepancy induced by the Cox–Merz rule. In this sense, dynamic measurements for viscosity can be considered when shear viscosity cannot be measured.

5.3.3 Mathematical Modeling of the Rheology and Related Numerical Analysis

As presented in the previous section, mucus exhibits a non-Newtonian behavior with shear-thinning effects. The previous section shows that the power-law is a good representation of the rheological behavior of the mucus. As presented in Table 5.3, the exponent of the shear rate is negative (since $n$ was identified below unity). From a mathematical and a computational point of view, this can be critical because the viscosity can tend toward infinity when the shear rate tends toward zero, in particular when the flow is at rest (at any given time and given point) or when any infinitely small fluid volume undergoes a solid-like displacement (i.e. with no strain).

In order to avoid the singularity that arises at rest, a limitation of the viscosity at low shear rates is introduced. Furthermore, the aqueous polymer mixture cannot be less viscous than its solvent, which means that at high shear rate, the decreasing viscosity has a lower bound at the PCL viscosity (Newtonian with viscosity equal to that of water at 20°C). Among the different constitutive laws, the Carreau model is interesting as it is a power-law with two Newtonian limits:

$$
\eta(\dot{\gamma}) = \eta_{\text{min}} + (\eta_{\text{max}} - \eta_{\text{min}}) \left[1 + (\beta \dot{\gamma})^\nu \right]^{\frac{n-1}{2}}.
$$

(5.24)
The transition between the plateau with viscosity \( \eta_{\text{max}} \), and the power-law behavior occurs at \( \dot{\gamma} = 1/\beta \). So \( \beta \) can be used for smoothing and avoiding the singularity at rest, and therefore can be chosen as large as wanted. The parameter \( \eta_{\text{max}} \) is then calculated from the power-law parameters and the \( \beta \)-value. In our simulations, \( \beta \) is set to 4000 s\(^{-1}\), this corresponds to a shear rate transition lower than the lowest shear rates explored in mucus rheometry. This \( \beta \) value was sufficient to ensure that the results do not depend on the regularization process. On the other hand, viscosity is limited to values higher than \( \eta_{\text{min}} \) (water viscosity here) for high shear rates.

Furthermore, the previous non-Newtonian results are given for the mucus layer, and the periciliary fluid layer remains a Newtonian fluid whose viscosity is equal to that of water. Hence it is necessary to model viscosity as a function of \( \alpha \): with a transition from the Newtonian periciliary fluid when \( \alpha = 0 \) to the non-Newtonian Carreau model when \( \alpha = 1 \).

From a numerical point of view, the only difference is the definition of the viscosity which is now a function of the velocity, through the shear rate \( \dot{\gamma}(u) = (2D : D)^{1/2} \). The problem to solve is the same as (5.11), except for the second equation which is replaced by an \( \alpha \)-dependent rheological constitutive equation (Chatelin et al., submitted for publication):

\[
\eta(\dot{\gamma}) = \Phi(u, \alpha) = \eta_{\text{PCF}} + (\eta_{0}(\alpha) - \eta_{\text{PCF}})[1 + (\beta \dot{\gamma})^2]^{(n(\alpha)-1)/2}
\]

where

\[
\eta_{0}(\alpha) = \eta_{\text{PCF}} \left( \frac{k_{\text{ML}}}{\eta_{\text{PCF}}} \right)^{\alpha} \beta^{\alpha(1-n_{\text{ML}})} \text{ and } n(\alpha) = \alpha \cdot n_{\text{ML}} + (1 - \alpha).
\]

As mentioned previously, these equations state that the viscosity of the mucus cannot be smaller than the periciliary fluid viscosity, even when the shear rate tends toward infinity. They are introduced in Eq. (5.2), which reads:

\[
-\text{div}(2\eta(\dot{\gamma})D) + \frac{1}{\varepsilon} \eta(\dot{\gamma}) \dot{\varepsilon}(u - \bar{u}) = f - \nabla p \text{ and } \text{div } u = 0 \text{ in } \Omega
\]

with the adequate boundary and initial conditions, coupled to the model followed by \( \alpha \), either the transport equation (5.13) or the stratified law displayed on Fig. 5.3.

Hence the computational algorithm presented in Section 5.2.2 is slightly modified. Indeed, the computation of \( \alpha \) remains identical whereas a non-linearity appears in the conservation of momentum equation. This non-linearity is treated “explicitly” in the same fixed point as the projection iterations. Hence the only modification is in the step 2b of this algorithm (see Section 5.2.2.3) which becomes a multi-criteria fixed-point algorithm. Denoting \( \eta_i = \Phi(u_i, \alpha) \), the new step 2b reads:

\[
-\eta_i \Delta u_{i+1} + \frac{1}{\varepsilon} \eta_i (u_i^{*} - \nabla \xi_i - \overline{\pi}(t)) = f + (2D(u_i) + \text{div } u_i^{*}) \nabla \eta_i.
\]

The rest of the algorithm presented in Section 5.2.2 remains unchanged. In particular, the SMW approach (see Section 5.2.2.4) is still applicable to compute (5.28) as well as the operator splitting and the Lagrangian computation of the convection of the mucin’s ratio.
5.3.4 Medical Outcomes

Regarding the Newtonian case, a parametric study was performed to investigate the influence of the rheological parameters on the mucociliary clearance efficiency (Chatelin et al., submitted for publication). The same quantity, defined by Eq. (5.22), is computed for this discussion. In Fig. 5.12, the results are presented for 169 \((n_{\text{ML}}, k_{\text{ML}})\) pairs using the \(\alpha\)-dependent rheological constitutive equations previously introduced. The mucus samples presented in Section 5.3, in Tables 5.2 and 5.3, are also reported on this phase cartography.

This figure clearly shows that the mucociliary transport does not evolve linearly with respect to the power-law coefficients \(n\) and \(k\). This is a direct consequence of the non-linearities of the modeling. In fact, it is not meaningful to discuss viscosity variations for shear-thinning fluids and these power-law coefficients are more significant. It is noteworthy to observe that the CF mucus generally takes lower consistency values \(k\) than healthy mucus samples, as well as lower power-law exponent \(n\) (except for the HL9).

Fig. 5.12 shows that a drop of 6–15% mobility may be observed for the CF mucus compared to the healthy samples. Even if this decrease is not spectacular, it is sufficient to trigger pathological infections such as *Pseudomonas aeruginosa* or *Burkholderia cepacia* (Eberl and Tümmler, 2004). Moreover, it is important to notice that even if the datum of HL9 (Serisier et al., 2009) is very different from other healthy mucus (Jeaneret-Grosjean et al., 1988; Zayas et al., 1990), the mean mucus velocity remains similar, and we can conclude that the associated mucociliary transports are comparable.

Fig. 5.12 demonstrates that the shear-thinning behavior has a very strong effect: when the power-law exponent decreases to 0.2 (which is the measured value for 5 samples), the mucociliary transport is highly deteriorated. This is counterintuitive, but it is the consequence of a higher velocity during the recovery stroke in the mucus layer, since the “apparent viscosity” decreases.
5.4 CONCLUDING REMARKS

From a clinical point of view, two types of CF mucus can be distinguished: stabilized and decompensated (with acute secondary infections). The mucus of same patient was sampled in both states: two days after hospitalization (CFM3a, decompensated) and after 17 days of treatment, combining antibiotics and steroids (CFM3b, stabilized). The computations presented in Fig. 5.12 clearly show an increase in the mucociliary transport for this patient, thus we conclude that the treatment was beneficial.

Finally, similarly to the Newtonian case, several computations were performed for different heights of periciliary fluid and still 169 values of \( (n_{ML}, k_{ML}) \) pairs – see Chatelin et al. (submitted for publication) for details. It was demonstrated that this periciliary fluid height has a very important effect on the mucociliary transport with up to a 21% decrease in mucus mean velocity. This parameter represents the effect of a variable CFTR deficiency (Thornton and Sheehan, 2004; Ehre et al., 2014) and it also shows the lack of mucociliary clearance in cystic fibrosis, allowing pathological proliferations.

This effect had been summarized in Fig. 5.13, where three groups are displayed:

- A group \( k = 1.5, n = 0.35 \), corresponding to a center of gravity of samples CFM3–5, 9, 10, which is moderately impacted by PCL rarefaction,
- A group \( k = 4, n = 0.25 \), corresponding to a center of gravity of samples CFM1, 6–8, 11, which is strongly impacted by PCL rarefaction, decreased up to \(-13\%\) compared to healthy cases, which means a \(-6\%\) due to lack of mucociliary clearance efficiency, made worse by \(-7\%\) due to PCL height decrease.
- A location \( k = 8, n = 0.15 \), corresponding to case CFL5 reported in Dawson et al. (2003), corresponding to a situation extremely impacted by PCL rarefaction, with a decrease of \(-23\%\) compared the healthy configuration.

The lung is resistant to damages following exposure to pathogens and more or less toxic chemical pollutants in inhaled air. Airway mucus represents the first line of defense. Inhaled particles (allergens, dust, micro-organisms, and cellular debris) trapped in the mucus gel are removed from the respiratory tract by the mucociliary clearance. Mucus cleans airways, as it flows from either the tracheobronchial tree or upper airways toward the pharynx, where it is swallowed (or expectorated).

Ciliary motions enable mucus propulsion in the human respiratory tract. The beating cycle of respiratory cilia is composed of 3 phases: a rest phase, a recovery stroke, and an effective stroke. During the recovery stroke, cilia (height 1–7 µm according to the location in the respiratory tract and investigators) sweep backward remaining close to the epithelium and immersed in the periciliary fluid without contact with mucus. Cilia of a patch of ciliated cells form a coordinated beating cluster (beating frequency 10–20 Hz). The cilium tips impact mucus during a short phase of its beating cycle. Cilia are more rigid in the forward power stroke and more flexible in recovery stroke in preparation for the next effective stroke.

The present mathematical model of airway surface liquid dynamics relies on a heterogeneous fluid that moves due to beating deformable cilia. The airway surface liquid nature evolves continuously in both composition and rheology from the periciliary fluid to the...
mucus. Within the near-wall fluid surrounding the deformable cilia, the mucin polymers released by the epithelium further hydrate. Once they reach a certain distance normal to the wetted surface of the respiratory epithelium, which is slightly lower than the cillum height, mucin polymers accumulate, engendering a mucin-concentrated phase. In this region, mucin polymers interweave and eventually cross-link. This evolving nature of the airway surface liquid affects its rheology. For example, for a airway surface liquid thickness of 12 µm and cillum height of 8 µm, the thickness of the periciliary fluid and mucus are equal to 7 and 5 µm, respectively.

The rheological behavior is associated with two parameters of interest, the fluid consistency index $k$ and the power law index $n$ that describes the difference between the two asymptotic behaviors ($\mu_0$ and $\mu_\infty$) and the slope of the shear rate–viscosity curve between these two asymptotic behaviors.
The numerical model is combined with an experimental study of the rheology of sputum from cystic fibrosis patients. The proposed robust numerical method handles variations in shear rate and viscosity of more than 5 orders of magnitude. It leads to a cartography that allows discussing major issues on defective mucociliary clearance observed in cystic fibrosis during periodic check-up of the disease in a steady clinical state (i.e., in the absence of infection exacerbation) for prognosis.

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References


REFERENCES


